Structural and functional analysis of “non-smelly” proteins

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Highlights

- Cysteine-depleted proteins are abundant in all domains of life
- Prokaryotes are significantly enriched in cysteine-depleted proteins compared to eukaryotes
- Only about 0.05% of proteins are depleted in aromatic residues and cysteine
- Proteins depleted in aromatic residues and cysteine have high levels of intrinsic disorder
- Organisms with higher levels of cysteine-depleted proteins have higher levels of the intrinsic disorder
- “Non-smelly” proteins are involved in translation, transcription, nucleosome assembly, protein folding, and transmembrane transport functions

Abstract

Cysteine and aromatic residues are major structure-promoting residues. We assessed the abundance, structural coverage, and functional characteristics of the “non-smelly” proteins; i.e., proteins that do

not contain cysteine residues (C-depleted) or cysteine and aromatic residues (CFYWH-depleted), across 817 proteomes from all domains of life. The analysis revealed that although these proteomes contained significant levels of the C-depleted proteins, with prokaryotes being significantly more enriched in such proteins than eukaryotes, the CFYWH-depleted proteins were relatively rare, accounting for about 0.05% of proteomes. Furthermore, CFYWH-depleted proteins were virtually never found in PDB. Depletion in cysteine and in aromatic residues was associated with the substantially increased intrinsic disorder levels across all domains of life. Archaeal and Eukaryotic organisms with higher levels of the C-depleted proteins were shown to have higher levels of the intrinsic disorder and lower levels of structural coverage. We also showed that the "non-smelly" proteins typically did not independently fold into monomeric structures, and instead they fold by interacting with nucleic acids as constituents of the ribosome and nucleosome complexes. They were shown to be involved in translation, transcription, nucleosome assembly, transmembrane transport, and protein folding functions, all of which are known to be associated with the intrinsic disorder. Our data suggested that, in general, structure of monomeric proteins is crucially dependent on the presence cysteine and aromatic residues.

Keywords: Intrinsically disordered proteins; cysteine-depleted proteins; nucleic acid binding proteins; proteins depleted in cysteine and aromatic residues; protein structure; protein function

Introduction

It is accepted now that intrinsically disordered proteins (IDPs) and hybrid proteins containing ordered domains and functionally important intrinsically disordered proteins regions (IDPRs) occupy a significant part of any proteome across all kingdoms of life and viruses [1-6], being especially abundant in eukaryotes [7,2]. Under physiological conditions, IDPs/IDPRs lack rigid 3D structure and therefore are typically not amenable to experimental structure determination by X-ray crystallography [8-10], which is by far the most commonly used technology to solve protein structures. As a result, they are considered as major constituents of the dark proteome [11,12,8]. While being disordered as a whole or in localized regions, these proteins have a number of important biological roles, especially in transcriptional and translational regulation, splicing, and signaling via cellular protein networks [13-15]. Furthermore, enhanced structural plasticity and exceptional spatiotemporal heterogeneity of IDPs/IDPRs define their mosaic structures, where different regions are disordered to different degrees. IDPs contain a multitude of potentially foldable, partially foldable, differently foldable or not foldable at all segments playing different roles in protein functionality [16,17], and even containing ordered regions that need to undergo order-to-disorder transition in order to make protein active [16,18,19]. In cellular protein-protein interaction networks, IDPs/IDPRs often play a role of hubs [20-24] that are engaged in promiscuous interactions and regulate the structural and functional integrity of these networks [25,26,15]. Furthermore, because of this binding promiscuity [27] and the ability to gain very different structures at binding to different partners [28], IDPs/IDPRs can "rewire" protein-protein interaction networks in response to environmental changes [29].

Systematic comparative analyses of amino acid sequences of ordered proteins and IDPs revealed the presence of numerous important differences [30-34,13]. For examples, extended IDPs/IDPRs from different kingdoms of life were shown to be rich in polar and charged amino acids and deficient in hydrophobic residues [30,35,34,36]. This also resulted in the elaboration of the concept of "order-promoting" (C, W, Y, I, F, V, L, H, T, and N) and "disorder-promoting" residues (A, G, D, M, K, R, S, Q, P, and E); i.e., residues more commonly found in ordered and disordered proteins/regions, respectively [37]. Because of the high relative enrichment of the amino acid sequences of ordered proteins and domains in cysteine, tryptophan, tyrosine, phenylalanine, and histidine, these residues are typically considered as strong order-promoting residues. Based on these observations, we hypothesized that structure and functionality of proteins can be noticeably dependent on the presence cysteine and aromatic residues in their amino acid sequences. One can argue that cysteine is important for protein structural stability only when another cysteine is present in the same chain, to enable disulfide bond formation. Observations below provide important evidence that this is not always correct. Intramolecular disulfide bonds are surely important stabilizing factors. For example, proteins and peptides containing cystine knot, which is a rotaxane-like structural motif containing three disulfide bridges, where a polypeptide region between two of those disulfides forms a loop, through which a third disulfide bond is threaded, are known to show a particularly high degree of structural stability [38,39]. There are also numerous examples in the literature, where the importance of intramolecular disulfide bonds for protein thermal stability was demonstrated (as systemized in [40]). As a result, introduction of additional disulfide bonds is considered as an attractive protein engineering strategy for generating proteins (e.g., antibodies) with enhanced conformational stability [41]. Furthermore, dysregulated cellular redox conditions leading to the alterations in the formation of native disulfide bonds are directly linked to various human diseases [42]. However, even a single cysteine contributes to protein structure stability, as it can be engaged in the intermolecular disulfide bond, or can exist as a free thiol and serve as a part of a protein catalytic site, or as a site of various posttranslational modifications (e.g., S-hydroxylation (S-OH), disulfide bond formation, phosphorylation, S-acylation, S-prenylation, protein splicing, N-acetylation, N-ADP-ribosylation, amidation, S-archaeol cysteine, cysteine sulfinic acid (-SO_2H) formation, methylation, N-myristoylation, nitrosylation, N-palmitoylation, S-palmitoylation, and S-glutathionylation) [43]. Furthermore, a single cysteine can be used for specific coordination of various ligands, e.g., metal ions. In fact, cysteine is known to show high affinity toward zinc ions (Zn^{2+}), and the resulting cysteine-Zn^{2+} complexes are important for protein structure, catalysis, and regulation [44], as seen in the CH_3-type zinc finger proteins [45,46] and in redox switches [44]. On the other hand, CD_3 motifs serves as a Mn^{2+} coordination group [47].

To check the validity of the hypothesis that the presence cysteine and aromatic residues is crucial for protein structure, we conducted here a comprehensive bioinformatics analysis of the "non-smelly" proteins; i.e., proteins depleted in cysteine and aromatic residues. Since cysteines are known to smell like rotten eggs [48] and since the side chains of W, Y, F, and H are aromatic (i.e., they contain aromatic ring systems, which are stable, cyclic, planar compounds with a ring of resonance bonds and which, unlike pure saturated hydrocarbons, might have specific odors/aroma), the proteins depleted in these residues are defined here as "non-smelly". In this study, we assembled a dataset of "non-smelly" proteins found in 817 complete proteomes, and also looked for such proteins in the Protein Data Bank (PDB) [49,50].

Materials and methods

Datasets

We analyzed a dataset of 817 complete proteomes, which are defined as collections of proteins encoded by the fully sequenced genome of a specific organism. We obtained these proteomes from the UniProt resource [51,52]. They cover 276,733 proteins from 64 Archaean organisms, 5,077,609 proteins from 552 Bacterial organisms and 4,208,817 proteins from 201 Eukaryotic organisms, for the total of 9,563,159 proteins. A complete list of the considered species is given in the Supplementary Materials. Table 1 provides a breakdown of these organisms into specific kingdoms/phyla.

We also examined proteins with solved structures collected from PDB. We limited our analysis to the wild-type protein chains that have the expression tags removed and that exclude peptides (chain length > 30 residues), and which cover majority of the corresponding full protein chain from UniProt (>60% coverage). We clustered the sequences of the considered PDB structures at 100% identity to remove duplicates. These steps ensure compatibility with the proteome-level analysis. We collected 99,461 chains that satisfy the aforementioned criteria (*PDB* dataset), as well as two of its subsets that include 50,301 chains that are in complex with nucleic acids (*PDB NA* dataset) and 7,413 monomers; i.e., single-chain structures that do not interact with other proteins and nucleic acids (*PDB monomer* dataset).

Computation of structural characteristics

We used computational methods to quantify content of putative intrinsic disorder (the fraction of residues that are predicted to be intrinsically disordered) and the current structural coverage (the fraction of proteins for which structure is available) on the whole-proteome scale. We evaluated the quality of the disorder content predictions using a large benchmark dataset that was recently used in [53,54] and which was originally published in [55]. We quantify the predictive quality by computing the mean absolute error (MAE) and Pearson correlation coefficient (PCC) between the disorder content predicted with the consensus and the native disorder content. The resulting MAE = 5.5% and PCC = 0.43, which suggests that the content predictions are relatively accurate and correlated with the native disorder content. Our consensus secures similar results for the subset of the benchmark proteins that have cysteine (MAE = 4.9% and PCC = 0.46) and that have above average cysteine content (MAE = 5.0% and PCC = 0.42).

Recent studies demonstrate that intrinsic disorder can be accurately predicted from protein sequences [55-59]. Furthermore, consensus-based approaches that combine outputs of several disorder predictors were shown to provide more accurate predictions when compared to single predictors [60-62]. For instance, consensus predictors more precisely quantify the disorder content (fraction of the disordered residues in a given protein sequence), reducing error by about 4% when compared to single predictors [61]. We applied a consensus of five complementary predictions produced by two popular tools, IUPred [63] and ESpritz [64]. They include results produced with two versions of the IUPred method, which

were designed to predict long (≥30 consecutive residues) and short disordered regions, and three versions of ESpritz that focus on the three types of annotations of disorder using: DisProt database [65,66], crystal structures from PDB, and NMR structures from PDB. These tools are characterized by competitive levels of predictive quality [57,55] and short runtime, which is critical to facilitate processing of over 9.5 million protein sequences. The consensus prediction of disorder requires that at least 3 out of 5 predictions indicate intrinsic disorder. The same consensus was applied in several related studies [2,67-72,8]. Our methodology is also similar to the consensus-derived putative disorder in MobiDB [73,74] and D^2P^2 [16] databases. We calculated the disorder content of a given dataset of proteins (e.g., proteome) which is defined as a fraction of residues predicted as disordered among all residues in that dataset.

We estimated the current structural coverage based on a computationally tractable approach proposed in [75] and recently used in [8,2,76]. For each protein we ran three rounds of PSI-BLAST [77] searches against the sequences of protein structures from PDB. A given proteins sequence that has >50 residues in length is annotated as having structure if it registers a hit in PDB with the E-value <0.001. In other words, structurally solved proteins are assumed to have at least one long segment of residues (representing at least one domain) that is sufficiently similar to a sequence of an already solved structure. The structural coverage of a proteome is defined as the fraction of the structurally solved sequences among all sequences in this proteome. Research shows that such PSI-BLAST-based estimates provide relatively accurate results. For instance, a similar PSI-BLAST-based approach failed to find templates (similar sequences that are structured) for only 3 out of 120 target proteins in CASP9 [78]. We recognize that there are more precise approaches to estimate structural coverage that are capable of finding remote homologs, such as I-TASSER [79,80], HHpred [81,82], and MODELLER [83,84]. However, these tools could not be scaled to the size of our dataset. To the best of our knowledge, the largest such attempt is the MODBASE resource that covers only 76 organisms [85]. We note that our estimates of the structural coverage are slightly underestimated by inadequately considering remote homologs. Nevertheless, this bias should be equally distributed across different proteomes, allowing us to perform comparative analyses between the corresponding organisms and domains of life.

Functional annotations using GO terms

We annotated protein functions and cellular locations that are associated with the proteins depleted in major order-promoting residues, such as cysteine, phenylalanine, tyrosine, tryptophan, and histidine. Such proteins were split into two groups, depleted in cysteine (C-depleted) and depleted in cysteine and aromatic residues: phenylalanine, tyrosine, tryptophan, and histidine (CFYWH-depleted). The corresponding functions/locations are significantly enriched in these proteins sets when compared with the proteins from the same domain of life. This analysis relies on the GO terms [86] collected from the UniProt resource. We excluded annotations with “potential”, “probable” and “by similarity” qualifiers that are generated using computer-predictions or indirect experimental evidence. We evaluated magnitude and statistical significance of the differences in the rates of occurrence of GO terms between the C-depleted (or CFYWH-depleted) proteins and a generic set of proteins in the same domain of life by following protocols defined in earlier related analyses [2,3,71]. This analysis was performed for each of the three types of GO terms: cellular components, biological processes and molecular functions. We

randomly selected half of the GO-annotated chains for a given C-/CFYWH-depleted protein set and compared them with the same number of chains/residues drawn at random from the same taxonomic domain. We ensured that proteins drawn from the same domain of life have the same chain length (with ± 10% tolerance), since the amount of intrinsic disorder, which indirectly affects protein function and location, is dependent on the chain length [87]. We repeated this 10 times and evaluated significance of the differences in the 10 sets of counts for the corresponding GO terms. If these measurements are normal, based on the Anderson-Darling test at 0.05 significance, then we applied the paired *t*-test (proteins sets are paired to match chain lengths) to evaluate the statistical significance of differences; otherwise we utilized the non-parametric paired Wilcoxon rank sum test. We considered only the differences with p-value<0.001 which also have large magnitude, i.e., the average enrichment in the C-/CFYWH-depleted protein set must be larger than 30%. We analyzed the enrichment of GO terms for the entire set of C/CFYWH-depleted proteins as well as for the subsets of fully disordered C-/CFYWH-depleted proteins.

Results and Discussion

Abundance of C-depleted and CFYWH-depleted proteins

We measured fraction of the C-depleted and the CFYWH-depleted proteins across the 817 proteomes and among the proteins in the three PDB-derived datasets. Table 1 summarizes these values across each domain of life and several larger kingdoms and phyla. About 28% proteins in Archaea, 19% in Bacteria and 8% in Eukaryota do not have cysteines. While we observe substantial differences in the abundance of the C-depleted proteins across the three domains of life while, these values are consistent across the kingdoms and phyla within each domain of life (Table 1). This observation suggests that this trend is broadly associated with domains of life. Only about 0.06% proteins in Archaea and 0.04% in Bacteria and Eukaryota do not have cysteine and aromatic residues. Table 1 shows that the abundance of the CFYWH-depleted proteins is similar across Bacteria and Eukaryota, with Archaea having somehow elevated levels of such proteins.

Table 1. Amount of cysteine-depleted (C-depleted) and cysteine and aromatic residues-depleted (CFYWH-depleted) proteins, disorder content and structural coverage for the considered 817 proteomes. We report median of these per-proteome measurements for the entire domains of life (in bold font) and several larger kingdoms/phyla. The domains of life and the phyla/kingdoms within each domain are arranged according to their overall fraction of C-depleted proteins.

Domain of life	Kingdom/phylum of life	Number of Species	Median (per proteome) fraction of C-depleted proteins [%]	Median (per proteome) fraction of CFYWH-depleted proteins [%]	Median (per proteome) disorder content [%]	Median (per proteome) structural coverage [%]
Archaea	**All**	**64**	**28.48**	**0.06**	**5.88**	**53.10**
	Crenarchaeota	17	34.05	0.06	3.00	53.56
	Other	4	24.71	0.11	4.45	55.85
	Euryarchaeata	43	19.81	0.07	5.10	53.96
Bacteria	**All**	**552**	**18.65**	**0.04**	**5.45**	**55.85**
	Firmicutes	76	22.62	0.04	4.60	60.07
	Actinobacteria	70	21.97	0.07	10.30	59.45
	Other	108	19.20	0.03	4.70	57.70
	Bacteroidetes	44	18.67	0.02	3.45	54.02
	Proteobacteria	254	16.99	0.04	5.80	61.37
Eukaryota	**All**	**201**	**7.87**	**0.04**	**19.70**	**47.70**
	Fungi	84	9.64	0.03	21.55	46.33
	Viridiplantae	15	7.37	0.04	16.40	45.84
	Other	31	5.40	0.04	16.80	41.71
	Metazoa	71	4.64	0.08	19.50	62.78

Figure 1 summarizes the distribution of the per-proteome abundance of the C-/CFYWH-depleted proteins for the three domains of life. The numbers of the C-depleted proteins vary significantly between Archaea, Bacteria and Eukaryota (p-values < 0.0001; **Figure 1A**) while the numbers of the CFYWH-depleted proteins are not significantly different (p-values ≥ 0.01; **Figure 1B**). Our analysis has revealed that prokaryotes harbor significantly larger numbers of the C-depleted proteins compared to eukaryotes. Furthermore, we compared these rates with the corresponding rates for the proteins with known structures collected from PDB. About 35% of proteins in the PDB dataset are depleted in cysteine (**Figure 1A**), while only two proteins (0.002%) are depleted in cysteine and aromatic residues (**Figure 1B**). The relatively high rate of the C-depleted proteins in PDB can be explained by two observations: about 2/3 of the PDB dataset is composed of the prokaryotic proteins; and because 51% of the proteins in this dataset were solved in complex with nucleic acids (PDB NA dataset). The effect of the latter factor is supported by our empirical finding that about 50% of the proteins in the PDB NA dataset are depleted in cysteine, which is a substantial enrichment particularly when compared to the PDB monomer dataset that has only about 19% of the C-depleted proteins (**Figure 1A**). We also emphasize the lack of the CFYWH-depleted proteins in PDB (**Figure 1B**). We found only two of them overall, with none in the PDB NA dataset and only one among the monomers. Importantly, the levels of the presence of the CFYWH-depleted proteins in PDB are substantially lower when compared to the rates of the CFYWH-depleted proteins in whole proteomes; i.e., 0.002% in PDB *vs*. 0.04% in Eukaryotic and Bacterial proteomes (20-fold decrease) and 0.06% in Archaean proteomes (30-fold decrease).

Altogether, these results demonstrate that the C-depleted proteins are significantly enriched in prokaryotes compared to eukaryotes and that they are often involved in protein-nucleic acids interactions and relatively rarely fold into monomer structures. On the other hand, the CFYWH-depleted proteins are equally abundant across the three domains of life and virtually never found in PDB. The latter suggest that they are hard to solve structurally.

CFYWH- and FYWH-depleted proteins in the PDB dataset and their intrinsic disorder status

Our search for the CFYWH-depleted proteins in the PDB dataset produced only two hits, a deletion mutant of the transcarboxylase biotin carrier subunit (also known as biotin carboxyl carrier protein, BCCP) from *Propionibacterium freudenreichii subsp. shermanii* (PDB ID: 1O78) and a molybdenum-pterin-binding protein 2 (molbindin-2 or MopII) from *Clostridium pasteurianum* (PDB ID: 1GUT), Functionally, BCCP serves as a carrier subunit of the transcarboxylase, which is a biotin-dependent 1200 kDa multi-subunit enzyme composed of 30 separable polypeptides [88]. Here, BCCP functions as a carboxyl group carrier to which biotin is covalently attached at Lys89. BCCP also binds the other two subunits of transcarboxylase to assist in the overall assembly of the enzyme [89]. NMR solution structure analysis revealed that the BCCP C-terminal domain (residues 51−123) is characterized by a compact β-sandwich structure, whereas the N-terminal region of the protein (residues 1-50) is disordered and does not have detectable structure [90]. **Figure 2A** represents the NMR solution structure of a CFYWH-depleted 10-48 deletion mutant (residues 1–9/49–123) of BCCP and shows that this protein contains six anti-parallel β-strands forming β-sandwich and a rather disordered N-terminal region. Furthermore, high flexibility was also detected at the C-terminal ‘β-finger’ segment of this deletion

mutant that contains the Lys89 biotinylation site [91]. The second CFYWH-depleted protein in PDB, molbindin-2, is a bacterial protein that serves as an intracellular storage facility for molybdate. **Figure 2B** shows that this protein exists as a hexamer assembled as a trimer of dimers and binds up to eight molybdate ions with high affinity [92]. A protomer of this protein has a twisted antiparallel β-sheet structure formed by five β-strands [92] (see **Figure 2C**).

Our analysis showed that the number of proteins in the PDB dataset that are depleted in the aromatic residues (FYWH-depleted) is also very low. In fact, we found only 7 such proteins, which, in addition to the aforementioned BCCP and molbindin-2 were a ribosomal protein S33 from *Trypanosoma brucei brucei (strain 927/4 GUTat10.1)* (which is a part of the bacterial ribosome, high resolution structure of which was solved by cryo-electron microscopy, PDB ID: 4V8M-AZ, see **Figure 2D**), a bacterial ribosomal protein S28E from *Methanobacterium thermoautotrophicum* (PDB ID: 1NE3, see **Figure 2E**), an eukaryotic 40S ribosomal protein rpS28e from *Tetrahymena thermophila* (PDB ID: 4V5O-A1/B1 and PDB ID: 4BTS-A1/B1/C1/D1, see **Figure 2F**), the pulmonary surfactant-associated polypeptide C (SP-C, PDB ID: 1SPF, see **Figure 2G**), and rat metallothionein-2 (PDB ID: 4MT2, see **Figure 2H**). Three of these FYWH-depleted proteins are ribosomal proteins, with the solution NMR structure of one of which (S28E) being solved at pH 4.5, and with two others (S33 and rpS28e) being a part of the ribosomal subunit). One of them (metallothionein-2) is a metal-binding protein that does not have any regular secondary structure elements and whose 3D structure is stabilized by homodimerization and coordination of five cadmium ions, two zinc ions, and one sodium ion [93]. The last one is a membrane-embedded protein (SP-C), whose structure in apolar solvent (a mixed solvent of C_2H_3Cl/C_2H_3OH/ 1 M HCl 32:64:5 (v/v)) was solved by NMR [94].

Since all CFYWH- and FYWH-depleted proteins in the PDB dataset are rather small and are characterized by strong amino acid biases (for example, metallothionein-2 possesses extremely high content of cysteine residues (32.8%)), next, we analyzed their intrinsic disorder predispositions using a set of commonly used per-residue disorder predictors, such as PONDR® VLXT [32], PONDR® VL3 [95], PONDR® VSL2 [96], IUPred_short [97] (yellow curve), IUPred_long [97], and PONDR® FIT [98]. **Figure 3** indicates that many of these proteins are predicted to have high levels of intrinsic disorder. In fact, according to their mean disorder predisposition, they can be ranged as follows: S33 (0.60±0.16) > rpS28e (0.52±0.17) > BCCP (0.47±0.14) = metallothionein-2 (0.47±0.44) > S28E (0.46±0.15) > molbindin (0.33±0.17) > SP-C (0.16±0.15). Low level of intrinsic disorder in SP-C was expected, since this is a transmembrane protein characterized by the high content of hydrophobic, order-promoting residues. **Figure 3** also shows that although, generally, the outputs of the predictors used in this study agree with each other, the disorder profile generated for metallothionein-2 reflects noticeable "confusion", where PONDR® VL3, PONDR® VSL2, and PONDR® FIT predicted this protein to be completely disordered, whereas IUPred_short and IUPred_long suggested that the metallothionein-2 is absolutely ordered. This discrepancy is defined by the highly biased amino acid sequence of this protein, which does not have aromatic residues, being instead heavily enriched in cysteine residues (20 of its 61 residues (32.8%) are cysteines).

One can argue that CWYFH-depleted proteins could contain a higher number of other (non-WYFH) hydrophobic amino acids and still be folded. Unfortunately, the amount of currently available data related to such proteins is not sufficient for conducting reliable statistical analysis to check this hypothesis. In fact, almost complete lack of the non-smelly proteins in PDB, which has only two

CWYFH-depleted and seven WYFH-depleted proteins, serves as an important indication that unique (foldable) protein structure requires cysteines and aromatic residues. Composition profiler-based [34] comparison of the amino acid compositions of two CWYFH-depleted proteins, BCCP and molbindin-2, with the amino acid compositions of globular proteins in PDB revealed that these non-smelly proteins are significantly enriched in valines (p-value<0.05). Extending this analysis to all seven WYFH-depleted proteins showed that they are significantly enriched in valines and methionines. However, the levels of other hydrophobic residues (leucines and isoleucines) were not significantly increased. These data are insufficient for making unambiguous conclusion on the presence of the compensatory increase in the number of non-CWYFH hydrophobic amino acids in the non-smelly proteins. Generally, since hydrophobic residues are order-promoting [37], one would expect that if such compensation would take place, then the resulting WYFH-depleted proteins with the increased content of non-WYFH hydrophobic residues would still be mostly ordered. However, we are showing here that proteins without CWYFH are more disordered than proteins with CWYFH (see below). This indicates that the proposed compensation is not globally observed. Of course there could be some exceptions from the rule, but there is no such compensation, in general. Furthermore, there is a logical limit on how many hydrophobic residues one can put into a sequence that can fold into a soluble structure (there is the surface to volume ratio limiting the number of hydrophobic groups that can be protected from water by a surface layer of hydrophilic residues upon formation of a globular structure).

In summary, the number of the CFYWH- and FYWH-depleted proteins in PDB is vanishingly small. Despite being structurally characterized, these proteins typically (with the noticeable exception of the pulmonary surfactant-associated polypeptide C, which is a highly hydrophobic, membrane binding protein) have rather content of intrinsic disorder. None of these proteins are enzymes. They are either oligomeric metal-binding proteins, or ribosomal proteins engaged in interaction with ribosomal RNA and other ribosomal proteins, or parts of protein complexes, or transmembrane proteins. In other words, none of these seven protein exist as a non-interacting monomer, suggesting that their structure is stabilized by interaction with binding partners. Therefore, it is safe to conclude that stable monomeric protein structure requires inclusion of cysteine and aromatic residues.

C- and CFYWH-depleted proteins are enriched in intrinsic disorder

The empirical observation that C-/CFYWH-depleted proteins are relatively rare in PDB suggests that they could be intrinsically disordered [99,8,10]. We tested this hypothesis utilizing accurate putative annotations of disorder. **Figure 4A** compares the putative disorder content (% of disordered residues) in all complete proteomes with the putative disorder content in the C-depleted and the CFYWH-depleted proteins for each domains of life. We found that proteins depleted in cysteine have relatively high disorder content at 7.8% in Archaea, 11.0% in Bacteria and 44.4% in Eukaryota. These are substantially higher amounts when compared to the corresponding complete proteomes. The increases relative to the complete proteomes range between (7.8-5.7)/5.7 = 37% in Achaea and (44.4-20.2)/20.2 = 120% in Eukaryota. The amounts of the putative intrinsic disorder are event higher among the CFYWH-depleted proteins, with 40.1% disorder content in Archaea, 60.9% in Bacteria, and over 80% in Eukaryota. When compared to the proteome-level disorder content, this corresponds to the relative increases by 604%, 867%, and 312%, respectively. **Figure 4B** compares fractions of the fully disordered proteins between the complete proteomes and the C-depleted and CFYWH-depleted protein datasets. The enrichment in

the number of fully disordered protein is even more substantial than for the disorder content. About 0.2% of all proteins *vs.* 0.7% of the C-depleted proteins in Archaea are fully disordered (250% increase), 0.4% *vs.* 1.4% in Bacteria (250% increase), and 1.0% vs. 8.2% in Eukaryota (820% increase). The corresponding increases when comparing the whole proteome-level amounts with the subset of the CFYWH-depleted proteins are approximately 8900% in Archaea and Bacteria and 6700% in Eukaryota. These results clearly demonstrate that the depletion in cysteine and in aromatic residues is associated with substantially elevated levels of intrinsic disorder across all domains of life.

We further analyzed proteome-level relation between the disorder content and the abundance of the C-depleted proteins, see **Figure 5A**. We did not pursue this analysis for the CFYWH-depleted proteins since their numbers are small relative to the proteome sizes (**Figure 1A**), and, therefore, they do not make sufficient impact on the proteome-level measurements. **Figure 5A** reveals a slight increase in the disorder content for organisms with high levels of C-depleted proteins; i.e., the linear fit is sloped upwards and the Pearson correlation coefficients (PCCs) are positive consistently across the three domains of life. This is in agreement with the domain-level increase in the intrinsic disorder for the C-depleted proteins shown in **Figure 4A**. We investigated whether this trends correlates with the current levels of structural coverage (% of proteins with at least partially known structures). **Figure 5B** shows relation between the current structural coverage and the amount of the C-depleted proteins for the three domains of life. We observe modest correlations for Archaea (PCC = -0.34) and Eukaryota (PCC = -0.43), and no correlation for Bacteria (PCC = 0.03). This suggests that archaeal and eukaryotic organisms with higher levels of the C-depleted proteins are characterized by lower levels of structural coverage. **Table 1** reveals that in case of the Eukaryotes this is driven by the high structural coverage and low fraction of the C-depleted proteins in Metazoa. This, in turn, is related to a strong taxonomic bias in the PDB, where 44% of protein structures (61,323 out of the total of 138,194) are from metazoan organisms, in spite of the fact that only 10% of currently sequenced proteins (13,484,303 out of 134,315,728 in UniProt) are from this kingdom of life. One possible explanation for the lack of the correlation in Bacteria is that these proteins have high propensity for crystallization, particularly in contrast to the eukaryotic proteins [76]. This has substantial influence since X-ray crystallography is the single biggest contributor to the protein structure determination efforts [100]; i.e., 90.3% (124,770 out of 138,194) protein structures in PDB were solved using X-ray crystallography. The visible decline in the structural coverage for Archaean proteins, which also have high propensity for crystallization [76], is likely a result of the significantly higher amount of the C-depleted proteins (**Figure 1A**), when compared to the Bacterial proteins. Overall, our empirical analysis reveals that archaeal and eukaryotic organisms with higher levels of the C-depleted proteins are characterized by higher levels of the intrinsic disorder and lower levels of the structural coverage.

Functional analysis of C-depleted and CFYWH-depleted proteins

Figure 6 lists cellular location and functions that are enriched among the C-depleted proteins. The analysis is broken into three types of annotations: cellular components (at the top of the figure), molecular functions (in the middle of the figure) and biological processes (at the bottom of the figure) and performed separately for each domain of life. The C-depleted proteins in Archaea and Bacteria are primarily localized in membranes and ribosome while in Eukaryota they are also found in the nucleosome and nucleolus. These subcellular locations point to a high likelihood that C-depleted

proteins are involved in the protein-RNA and protein-DNA interactions. Molecular functions listed in **Figure 6** reveal that indeed they interact with the rRNAs and, in Eukaryotes, with DNA, while also being involved in the transporter and motor functions. Since the C-depleted proteins are enriched in the intrinsic disorder, these observations are further supported by literature that suggests that IDPs and IDPRs play key roles in the protein-nucleic acids interactions [71,101,69,102,68,103,104,30,105,32,106,107]. The biological processes associated with the C-depleted proteins are consistent with the aforementioned observations, and they cover translation, protein folding, nucleosome assembly, and protein transport. The subset of fully disordered C-depleted proteins (FD lines in **Figure 6**) can be found in ribosome, nucleosome, and, in Eukaryotes, in the chromatin. These proteins implement translation in Archaea and Bacteria, and they also carry out several other functions in Eukaryota, such as response to stress and spermatogenesis. Overall, our analysis reveals that protein-nucleic acids interactions underlie cellular functions and locations of the C-depleted proteins.

Figure 7 summarizes the major subcellular locations and functions that are enriched in the CFYWH-depleted proteins. These proteins are primarily found in ribosome in Archaea and Bacteria, while in Eukaryota, they are also located in the nucleus, particularly in the chromatin, by being part of the nucleosome complex. This is in agreement with the observations that these proteins are significantly enriched in disorder (**Figure 4**), and that the nucleosome and ribosome complexes contain proteins enriched in disordered regions [71,69,102,68]. The molecular functions and processes associated with the CFYWH-depleted proteins involve RNA and DNA binding in the context of translation, nucleosome assembly, and transcription. This is again consistent with earlier studies that revealed that the high levels of intrinsic disorder represent one of the important characteristics of the nucleic acid binding proteins [71,101,69,102,68,103,104,30,105,32,106,107]. Furthermore, the spatially and temporally coordinated action of many macromolecular complexes and proteins containing functionally significant IDPRs represents an important means for the control of transcription [108]. The major stages of transcription include chromatin remodeling that regulates the global accessibility of promoter DNA, action of regulatory transcription factors, co-activators/co-repressors, and the basal transcription machinery, and at each of these stages, intrinsically disordered proteins or proteins with IDPRs play very important regulatory roles [108]. Next, we discuss the role of disordered nucleic acid-binding proteins in each of these stages.

Formation of the nucleosomes, which are the basic structural units of chromatin, represents the primary step in the DNA condensation that is strongly protein intrinsic disorder-dependent. Nucleosomes are formed via association of small, highly basic nuclear proteins, core histones, with DNA in a specific stoichiometry. The formed nucleosomes are condensed together via action of the linker histones. Comprehensive bioinformatics analyses of 2007 histones from 746 species revealed that all the members of the histone family are highly disordered and utilize disorder for various functions, such as heterodimerization, formation of higher order oligomers, interaction with DNA and other proteins, and posttranslational modifications [102]. Among nuclear proteins that bind to nucleosomes, alter the structure of chromatin, and affect transcription are the members of a high mobility group N (HMGN) protein family of highly disordered chromatin modifying proteins [109]. In addition to HMGNs, many other IDPs and proteins containing functionally important IDPRs, including various chromatin modifying enzymes, are involved in the regulation of DNA accessibility [108].

Among the most illustrative examples of IDPs related to the regulation of transcription (after the chromatin environment becomes accessible due to the actin of chromatin remodeling proteins) are transcription factors (TFs, which are also known as sequence-specific DNA-binding factors). TFs are multifunctional proteins which are crucial for the control of expression of specific genes and for the regulation of the gene activity in response to specific stimuli. They deliver their effects via binding to specific DNA sequences, recruiting the RNA polymerase to specific genes, controlling the transfer of genetic information from DNA to mRNA, and positively or negatively influencing the gene transcription either alone or in a complex with other proteins [110]. Generally, the modular structure of TFs includes one or more DNA-binding domains (DBDs) for recognition and binding of the specific DNA sequences adjacent to the genes that they regulate, and one or more transactivation domains for recognition of the co-activators and/or other transcription factors. Computational analysis of several TF datasets revealed that between 82.6% and 94.1% of TFs possess long IDPRs, with the degree of disorder being significantly higher in eukaryotic FTs in comparison with their prokaryotic counterparts [111]. TFs also contain high levels of disorder-based protein interaction sites, molecular recognition features (MoRFs) [111]. Intrinsic disorder is not distributed evenly within the sequences of TFs. In fact, although in general the DNA-binding domains are noticeably less disordered than the TF activation regions (or transactivator domains), the AT-hooks and basic regions of DNA-binding domains of TFs are highly disordered [111]. In human TFs, almost 50% of the entire sequences are occupied by IDPRs [112]. Intrinsic disorder of transactivator domains is used in communication of TFs with other regulatory transcriptional proteins and has an important role in orchestrating the transcriptional assemblies [108]. Based on the high prevalence and versatility of intrinsic disorder in eukaryotic TFs, it has been concluded that these proteins can be used as important illustrations of various aspects of intrinsic disorder-based functionality [113].

At the next stage of transcription, co-activators and co-repressors define a cross-talk between chromatin, transcription factors, and the basal transcription machinery. Some of the co-activators can be considered as scaffolds containing multiple transcription factor binding sites and thereby processing multiple transcriptional regulatory inputs. One of such co-activators, p300, is known to interact with over 50 proteins and possesses histone acetyltransferase activity. Another illustrative example of the importance of intrinsic disorder in the transcription regulation is the Mediator complex. This complex serves as an interface between gene-specific regulatory proteins and the general transcription machinery and it contains high levels of functional intrinsic disorder [114].

Similarly, many proteins related to translation (i.e., the process of ribosome-mediated biosynthesis of proteins from mRNA) are either intrinsically disordered or contain long IDPRs. For example, ribosomal proteins are considered as an important example of the exceptional functional versatility of the RNA-binding IDPs. Based on the comprehensive bioinformatics analyses of the 3,411 ribosomal proteins from 32 species it has been concluded that many ribosomal proteins are either intrinsically disordered as a whole or represent hybrids containing ordered and disordered domains, and that intrinsic disorder is absolutely crucial for their various functions [69]. In agreement with these observations, our analysis showed that three of seven FYWH-depleted proteins whose structure is present in PDB are ribosomal proteins.

Taken together, our analysis revealed that although proteins that do not contain cysteines constitute rather large fraction of the analyzed proteomes (content of such C-depleted proteins is ranging from 8%

proteins in Eukaryota to 19% in Bacteria and to 28% in Archaea), proteins that do not have cysteine and aromatic residues (CFYWH-depleted proteins) constitute only very minor fractions of 817 complete proteomes (about 0.06% proteins in Archaea and 0.04% proteins in Bacteria and Eukaryota). Archaeal and Eukaryotic organisms with higher levels of the C-depleted proteins are predicted to have higher intrinsic disorder levels and lower structural coverage levels, whereas CFYWH-depleted proteins across all domains of life are characterized by the substantially increased levels of intrinsic disorder. Functional analysis revealed that the "non-smelly" proteins are often involved in protein-nucleic acids interactions. They are rarely present as independently folded monomeric structures and often serve as parts of the ribosome and nucleosome complexes. They are also found in cellular membranes. These C- and CFYWH-depleted "non-smelly" proteins are involved in translation, transcription, nucleosome assembly, transmembrane transport, and protein folding functions, all of which are known to be associated with the intrinsic disorder.

Finally, described in this article general inability of the "non-smelly" proteins to fold into self-organizing monomeric structures provides support to the hypothesis on the highly disordered nature of the primordial proteins, which is based on an intriguing correlation between the evolutions of genetic code and protein structure [115-117]. In fact, it was pointed out that the "prebiotic set" of amino acids (i.e., a set of amino acids that were generated by various abiotic processes) likely included 10 of 20 modern amino acids, such as A, D, E, G, I, L, P, S, T, and V [118,119], many of which were disorder-promoting. Based on a combination of 40 different factors, Eduard Trifonov proposed the following temporal order of addition of the amino acids to the genetic code: G/A, V/D, P, S, E/L, T, R, N, K, Q, I, C, H, F, M, Y, W [120]. This sorting underscores the correlation between the appearance of early amino acids (such as G, D, E, P, and S) in the primordial soup and their disorder-promoting tendencies in IDPs. In contrast, it seems that the major order-promoting residues, such as C, W, Y, F, and H, have been added to the genetic code at later evolutionary stages [116,115]. In other words, primordial proteins were "non-smelly". Similar inferences were also made by Brooks *et al*. in their study on the amino acid composition of last universal ancestral genomes [121]. Also, it was pointed out that the emergence of the biosynthesis of aromatic amino acid enabled an early halophile-to-mesophile transition, emphasizing the potential role of aromatic residues in the adaptive spread of early life and suggesting a selective advantage for the incorporation of aromatic amino acids into the codon table [122]. Furthermore, the high prevalence of nucleic acid binding-related functions among the modern "non-smelly" proteins can be considered as a kind of functional fossil, since nucleic acid binding and RNA chaperoning were proposed to be the first functions of primordial polypeptides [123,124]. Such RNA chaperone activities of early proteins provided their carriers a significant selective advantage in the RNA world, where RNA, which is especially prone to misfolding [125,126], was used for both information storage and catalysis [127].

Conclusions

We report the results of a comprehensive bioinformatics analysis of the prevalence and functionality of the "non-smelly" proteins (i.e., proteins that do not contain cysteine and aromatic residues, C- and CFYWH-depleted proteins) among 9,563,159 proteins from the 817 complete proteomes, and among the 99,461 PDB proteins with known 3D structures. This analysis revealed that prokaryotes are significantly enriched in the C-depleted proteins compared to eukaryotes. In fact, 28% proteins in

Archaea and 19% in Bacteria vs. only 8% in Eukaryota do not have cysteines. In general, C-depleted proteins are often involved in protein-nucleic acids interactions and they relatively rarely fold into monomer structures. On the other hand, CFYWH-depleted proteins are rather rare, are equally distributed across the three domains of life, and are virtually never found in PDB. Only about 0.05% of proteins do not have cysteine and aromatic residues. Depletion in cysteine and in aromatic residues is associated with the substantially elevated levels of intrinsic disorder in proteins across all domains of life. Archaeal and Eukaryotic organisms with higher levels of the C-depleted proteins have higher levels of the intrinsic disorder and lower levels of structural coverage. The C- and CFYWH-depleted proteins are part of the ribosome and nucleosome complexes and are also found in cellular membranes. They are involved in translation, transcription, nucleosome assembly, transmembrane transport and protein folding functions, all of which are known to be associated with the intrinsic disorder.

In line with highly disordered nature of the "non-smelly" proteins, is an important observation that such proteins are highly underrepresented in PDB. As a matter of fact, there are only two CFYWH-depleted proteins and five FYWH-depleted proteins among the ten thousand proteins in the PDB datasets which were solved by X-ray crystallography, or NMR, or cryo-EM. Furthermore, only two of these proteins, deletion mutant of BCCP and ribosomal protein S28E, have structures that can be considered as a result of a spontaneous folding of a single polypeptide chain, whereas structures of the other "non-smelly" proteins are stabilized by binding of metal ions and self-oligomerization (metallothionein-2 and molbindin-2) or by inclusion into large ribonucleoprotein complexes (ribosomal proteins S33 and rpS28e), or by placing into the non-polar solvent (pulmonary surfactant-associated polypeptide C). These observations indicate that a self-foldable unique 3D-structure in a globular protein is crucially dependent on the presence of cysteine and aromatic residues in its amino acid sequence.

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Conflict of interest

The authors have declared no conflict of interest.

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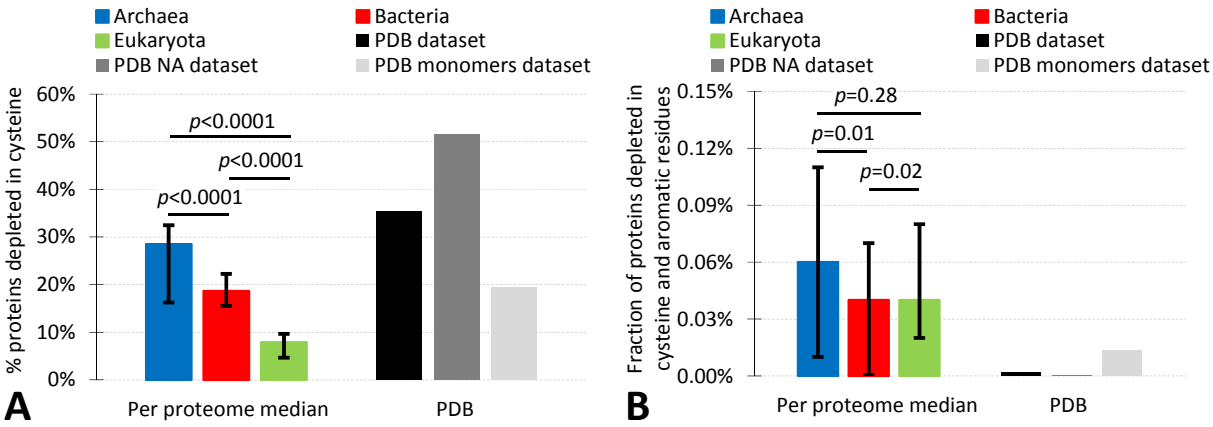

Figure 1. Abundance of C-depleted (panel A) and CFYWH-depleted proteins (panel B) in the three domains of life and among the structurally solved proteins from PDB. The blue, red and green bars show the median per-proteome fraction of C-/CFYWH-depleted proteins among the 64 Archaean, 552 Bacterial and 201 Eukaryotic organisms, respectively. The whiskers denote the first and third quartiles of these per-proteome fractions. Statistical significance of the differences for the per-proteome values between domains of life was assessed with the Wilcoxon test for unpaired data; distributions of the measured values are not normal. The black, dark gray and light gray bars show the fraction of the C-/CFYWH-depleted proteins among wild-type proteins chains from PDB (PDB dataset), wild-type PDB proteins that interact with nucleic acids (PDB NA dataset) and wild-type PDB monomers (PDB monomers dataset), respectively.

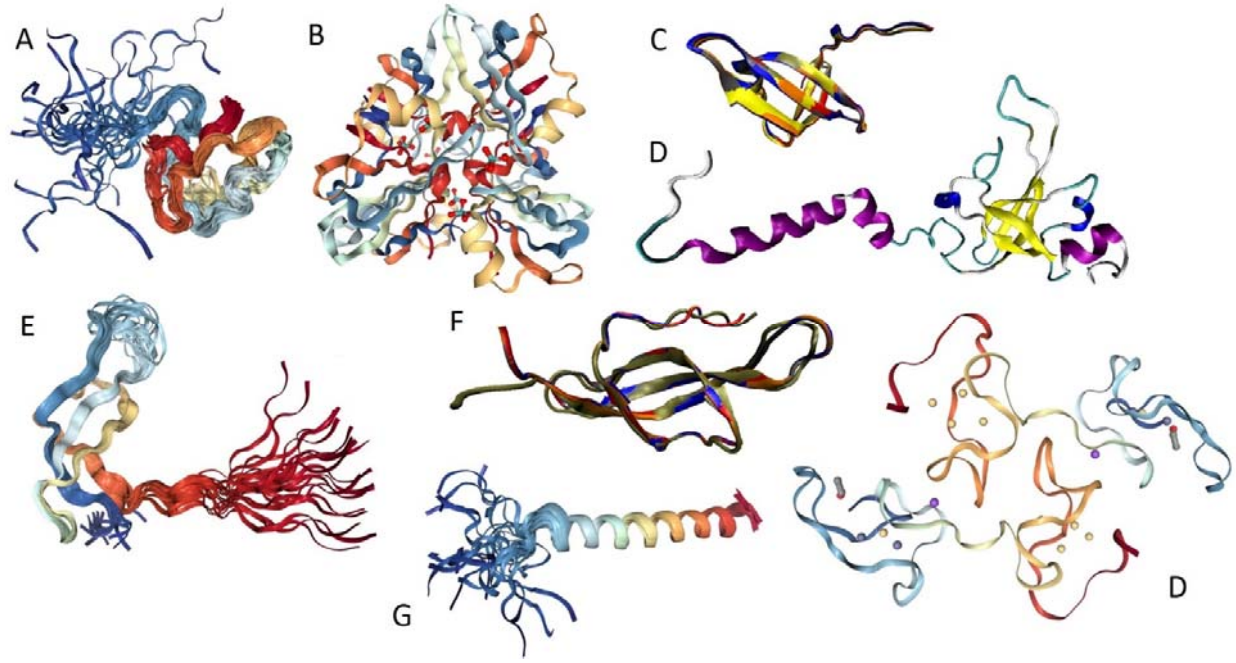

Figure 2. Structural characterization of the CFYWH- and FYWH-depleted proteins found in PDB: **A**. NMR solution structure of a CFYWH-depleted 10-48 deletion mutant (residues 1–9/49–123) of the transcarboxylase biotin carrier subunit (also known as biotin carboxyl carrier protein, BCCP) from *Propionibacterium freudenreichii subsp. shermanii* (PDB ID: 1O78); **B**. Crystal structure of the homohexameric molybdenum-pterin-binding protein 2 (molbindin-2 or MopII) from *Clostridium pasteurianum* (PDB ID: 1GUT); **C**. Aligned structures of the molbindin-2 protomers. Structures were aligned using a MultiProt server [128]. Plot was created using VMD platform [129]; **D**. High-resolution cryo-electron microscopy structure of a 40S ribosomal protein S33 from *Trypanosoma brucei brucei (strain 927/4 GUTat10.1).* Structure of this protein was extracted from of the cryo-EM structure of bacterial ribosome (PDB ID: 4V8M-AZ). Plot was created using VMD platform [129]; **E**. Solution NMR structure of a 30S bacterial ribosomal protein S28E from *Methanobacterium thermoautotrophicum* (PDB ID: 1NE3); **F**. Aligned structures of a eukaryotic 40S ribosomal protein rpS28e from *Tetrahymena thermophila* (PDB ID: 4V5O-A1/B1 and PDB ID: 4BTS-A1/B1/C1/D1). Corresponding structures were extracted from the crystal structures of the eukaryotic 40S ribosomal subunit in complex with initiation factor 1 (PDB ID: 4V5O-A1/B1) and the crystal structure of the eukaryotic 40S ribosomal subunit in complex with eIF1 and eIF1A (PDB ID: 4BTS-A1/B1/C1/D1). Structures were aligned using a MultiProt server [128]. Plot was created using VMD platform [129]; G. Solution NMR structure of the pulmonary surfactant-associated polypeptide C (SP-C) solved in apolar solvent (a mixed solvent of C_2H_3Cl/C_2H_3OH/ 1 M HCl 32:64:5 (v/v)) (PDB ID: 1SPF); and **H**. Crystal structure of the metal-bound dimer rat metallothionein-2 (PDB ID: 4MT2).

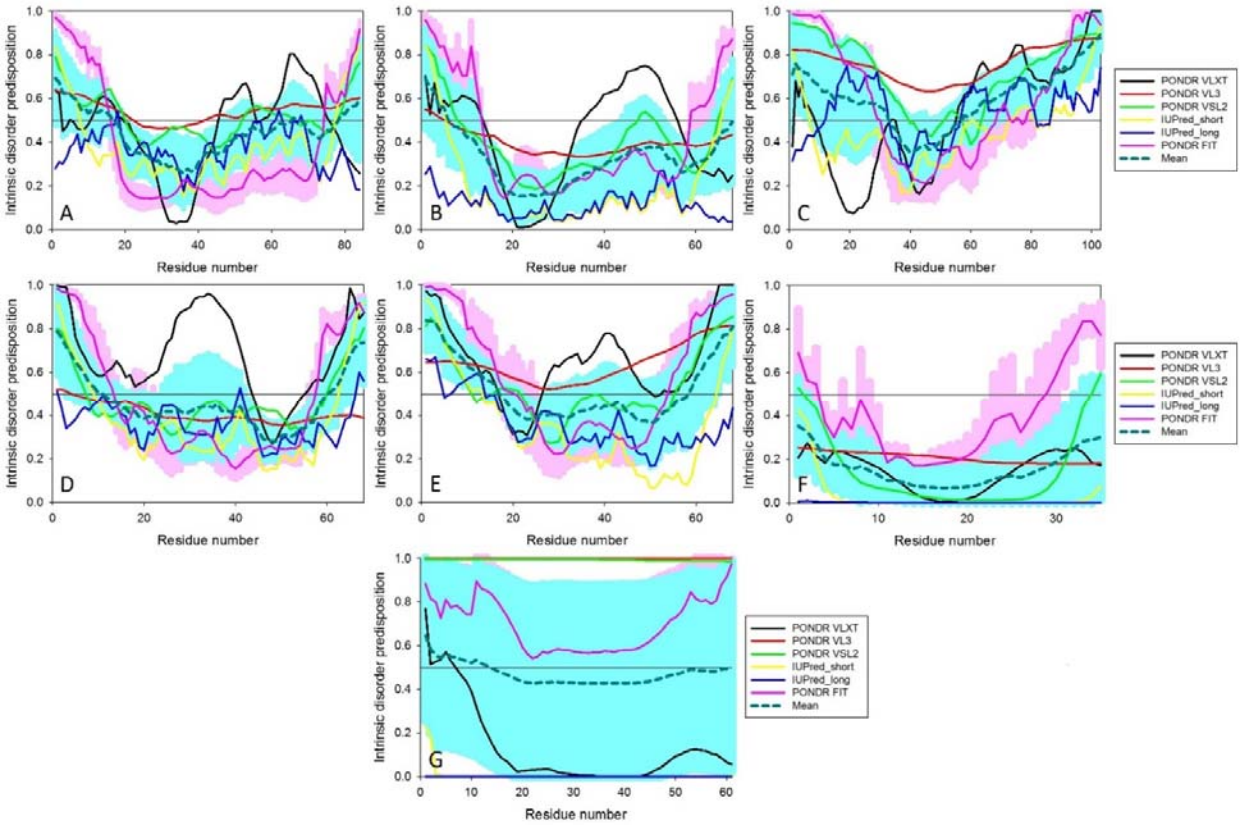

Figure 3. Multiparametric analysis of the intrinsic disorder predisposition of the CFYWH- and FYWH-depleted proteins found in PDB by several common predictors of intrinsic disorder: PONDR® VLXT [32] (black curves), PONDR® VL3 [95] (red curves), PONDR® VSL2 [96] (green curves), IUPred_short [97] (yellow curves), IUPred_long [97] (blue curves), and PONDR® FIT [98] (pink curves). Dark cyan dashed line shows the mean disorder propensity calculated by averaging disorder profiles of individual predictors. Light pink shadow around the PONDR® FIT shows error distribution for this predictor, whereas light cyan shadow around the mean disorder curve shows error distribution for evaluation of mean disorder. In these analyses, the predicted intrinsic disorder scores above 0.5 are considered to correspond to the disordered residues/regions, whereas regions with the disorder scores between 0.2 and 0.5 are considered flexible. Analyzed proteins were **A**. BCCP (residues 1–9/49–123 of UniProt ID: 02904); **B**. Molbindin-2 (UniProt ID: P08854); **C**. 40S ribosomal protein S33 from *Trypanosoma brucei* (UniProt ID: Q57U30); **D**. 30S ribosomal protein S28E from *Methanobacterium thermoautotrophicum* (UniProt ID: O26356); **E**. 40S ribosomal protein rpS28e from *Tetrahymena thermophila* (UniProt ID: Q234G5); **F**. pulmonary surfactant-associated polypeptide C (UniProt ID: P15785); and **G**. metallothionein-2 (UniProt ID: P04355).

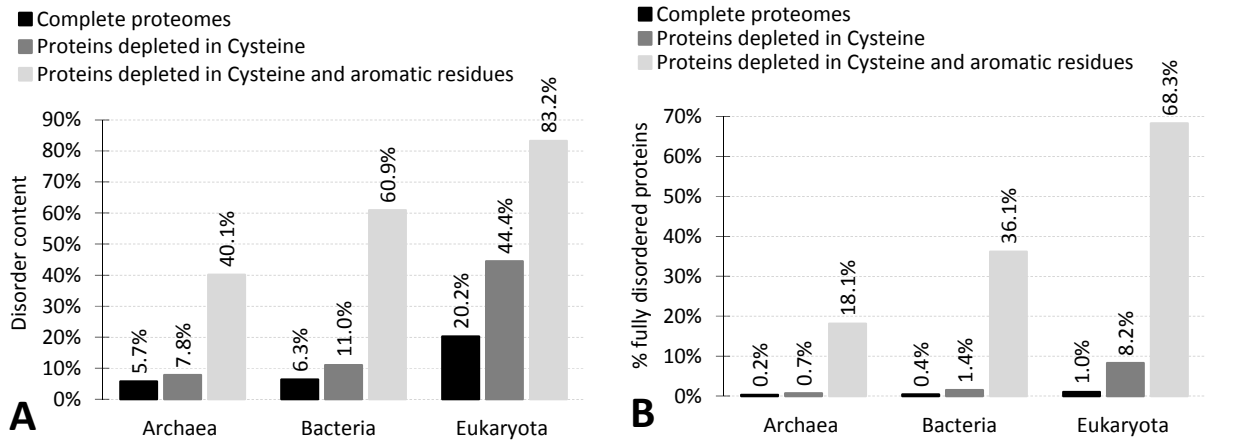

Figure 4. Comparison of the disorder content (panel A) and fraction of fully disordered proteins (panel B) between complete proteomes, C-depleted proteins and CFYWH-depleted proteins in the three domains of life.

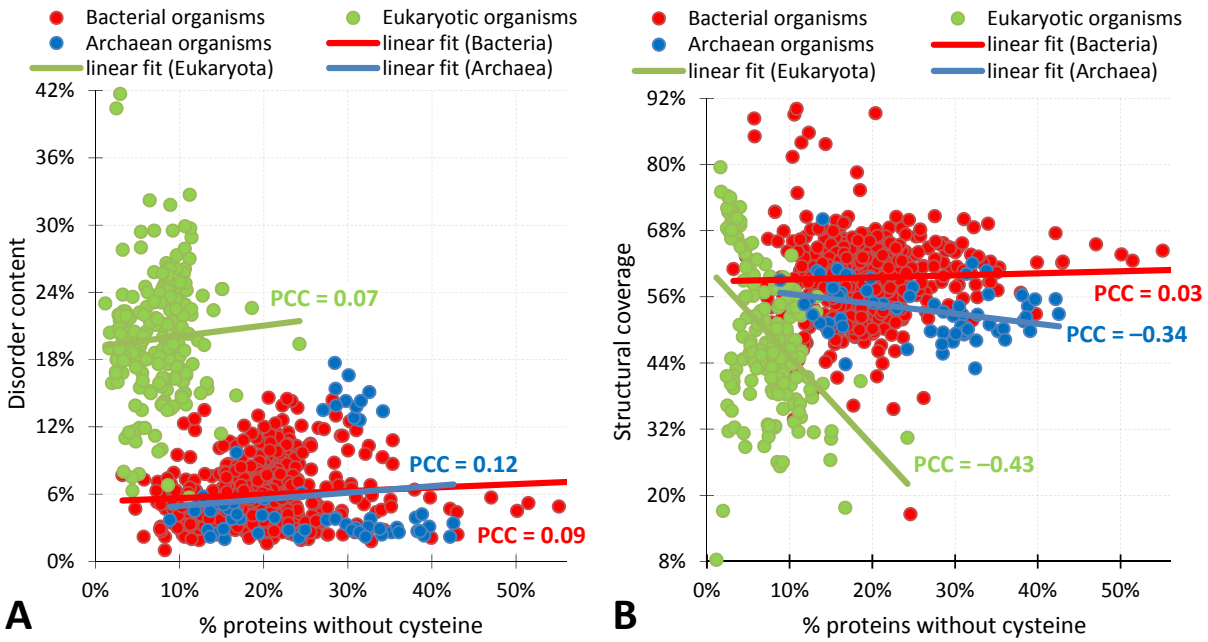

Figure 5. Relation between proteome-level abundance of the C-depleted proteins and structural coverage (panel A) and disorder content (panel B). Each point represents a single proteome. Lines represent linear fit into the data for a specific domain of life, which is accompanied with the corresponding values of the Person correlation coefficient (PCC).

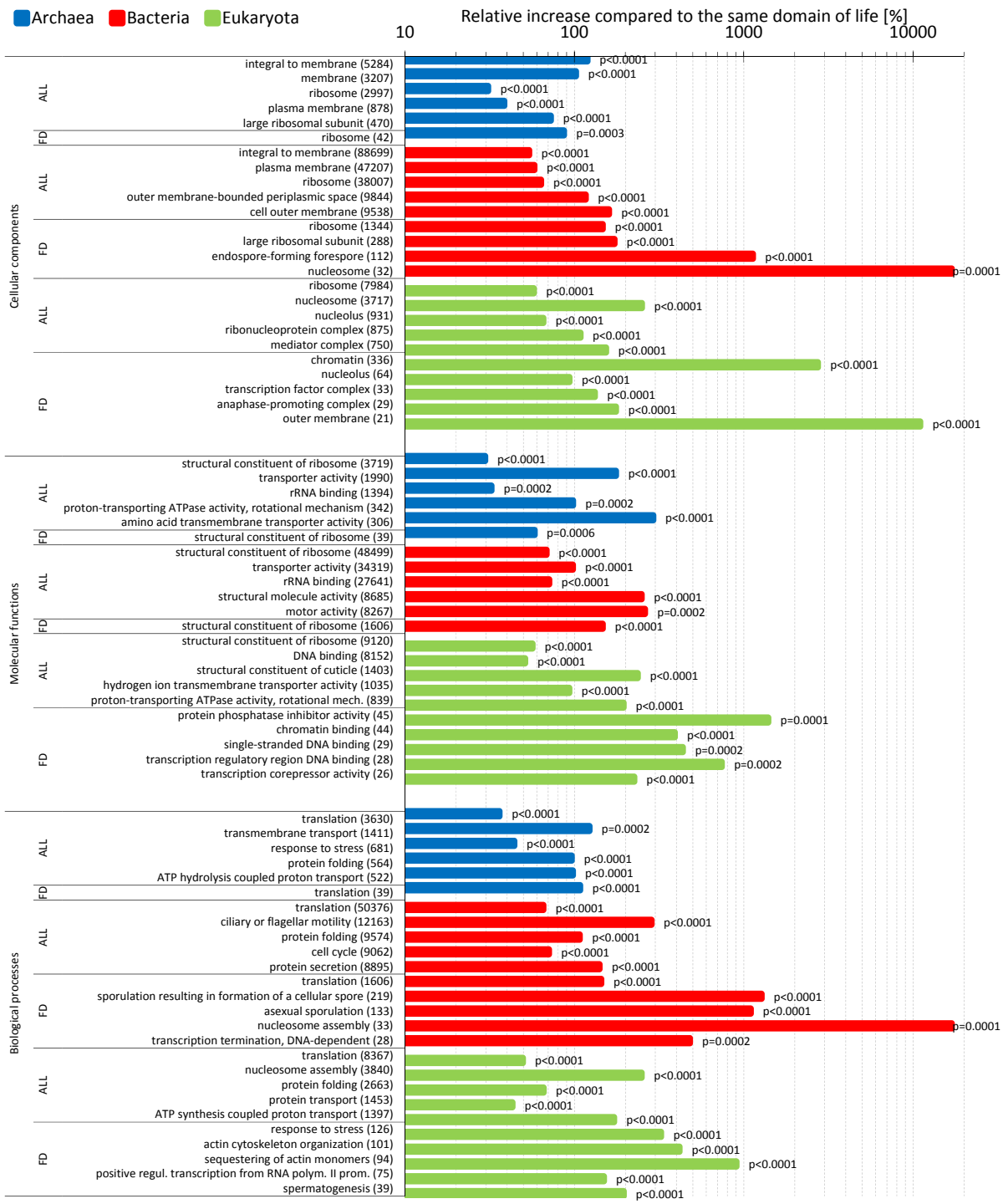

Figure 6. Cellular components (top of the figure), molecular functions (middle of the figure) and biological processes (bottom of the figure) that are significantly enriched in all C-depleted proteins (ALL lines) and among fully disordered C-depleted proteins (FD lines). The analysis is performed separately for eukaryotic species (green bars), bacterial species (red bars) archaeal species (blue bars). The y-axis lists five most frequent (the corresponding number of annotated proteins is shown inside the brackets) and significantly enriched functions/components (*p*-value < 0.001 and enrichment > 30%). The x-axis shows the enrichment measured as relative increase in frequency when compared to the size and chain length matched set of randomly chosen proteins from the same domain of life. Details of the calculation are explained in the Materials and Methods section. The functions/cellular components are sorted, within each group, by the number of annotated proteins.

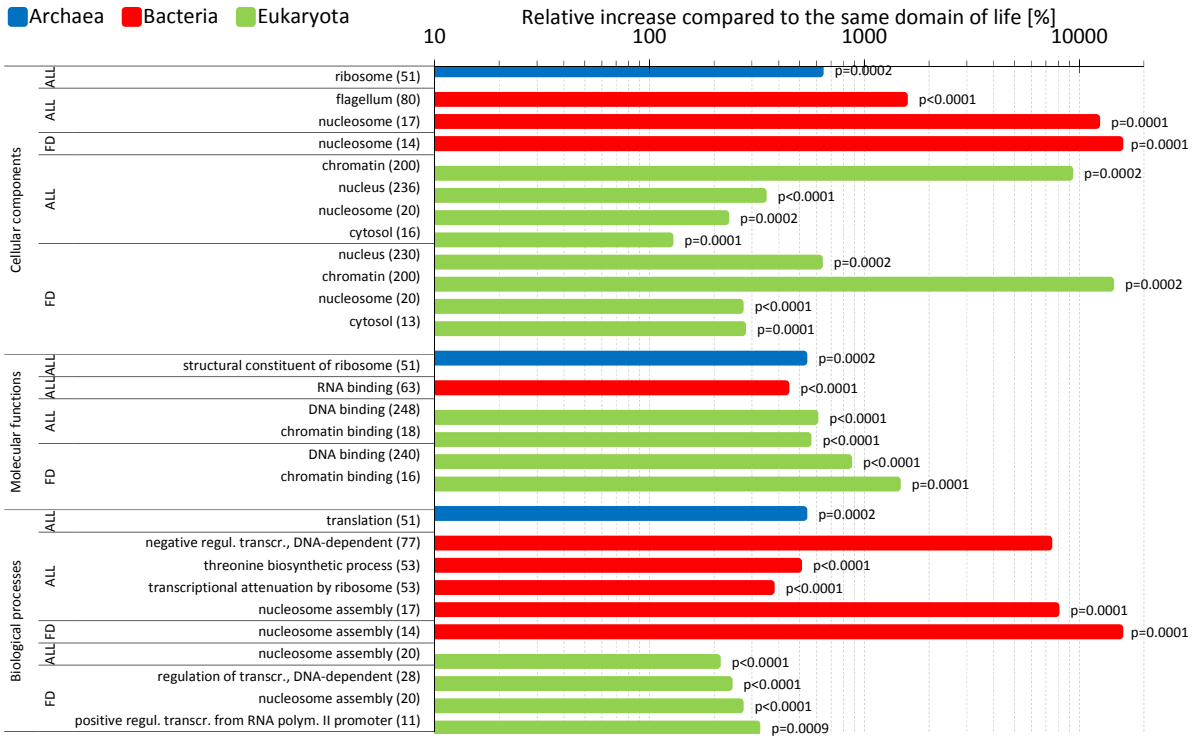

Figure 7. Cellular components (top of the figure), molecular functions (middle of the figure) and biological processes (bottom of the figure) that are significantly enriched in all CFYWH-depleted proteins (ALL lines) and among fully disordered CFYWH-depleted proteins (FD lines). The analysis is performed separately for eukaryotic species (green bars), bacterial species (red bars) archaeal species (blue bars). The y-axis lists five most frequent (the corresponding number of annotated proteins is shown inside the brackets) and significantly enriched functions/components (p-value < 0.001 and enrichment > 30%). The x-axis shows the enrichment measured as relative increase in frequency when compared to the size and chain length matched set of randomly chosen proteins from the same domain of life. Details of the calculation are explained in the Materials and Methods section. The functions/cellular components are sorted, within each group, by the number of annotated proteins.

Supplementary materials

The supplementary materials include the list and taxonomic classification of the 817 considered proteomes.

DomainOfLife	TaxonomicClassification
Archaea	Crenarchaeota Thermoprotei Acidianus
Archaea	Crenarchaeota Thermoprotei Acidilobus
Archaea	Crenarchaeota Thermoprotei Aeropyrum
Archaea	Crenarchaeota Thermoprotei Caldivirga
Archaea	Crenarchaeota Thermoprotei Desulfurococcus
Archaea	Crenarchaeota Thermoprotei Hyperthermus
Archaea	Crenarchaeota Thermoprotei Ignicoccus
Archaea	Crenarchaeota Thermoprotei Ignisphaera
Archaea	Crenarchaeota Thermoprotei Metallosphaera
Archaea	Crenarchaeota Thermoprotei Pyrobaculum
Archaea	Crenarchaeota Thermoprotei Pyrolobus
Archaea	Crenarchaeota Thermoprotei Staphylothermus
Archaea	Crenarchaeota Thermoprotei Sulfolobus
Archaea	Crenarchaeota Thermoprotei Thermofilum
Archaea	Crenarchaeota Thermoprotei Thermoproteus
Archaea	Crenarchaeota Thermoprotei Thermosphaera
Archaea	Crenarchaeota Thermoprotei Vulcanisaeta
Archaea	Euryarchaeota Aciduliprofundum
Archaea	Euryarchaeota Archaeoglobi Archaeoglobus
Archaea	Euryarchaeota Archaeoglobi Ferroglobus
Archaea	Euryarchaeota Halobacteria Halalkalicoccus
Archaea	Euryarchaeota Halobacteria Haloarcula
Archaea	Euryarchaeota Halobacteria Halobacterium
Archaea	Euryarchaeota Halobacteria Haloferax
Archaea	Euryarchaeota Halobacteria Halogeometricum
Archaea	Euryarchaeota Halobacteria Halomicrobium
Archaea	Euryarchaeota Halobacteria Halopiger
Archaea	Euryarchaeota Halobacteria Haloquadratum
Archaea	Euryarchaeota Halobacteria Halorhabdus
Archaea	Euryarchaeota Halobacteria Halorubrum
Archaea	Euryarchaeota Halobacteria Haloterrigena
Archaea	Euryarchaeota Halobacteria Natrialba
Archaea	Euryarchaeota Halobacteria Natronomonas
Archaea	Euryarchaeota Methanobacteria Methanobacterium
Archaea	Euryarchaeota Methanobacteria Methanobrevibacter
Archaea	Euryarchaeota Methanobacteria Methanosphaera
Archaea	Euryarchaeota Methanobacteria Methanothermobacter
Archaea	Euryarchaeota Methanobacteria Methanothermus
Archaea	Euryarchaeota Methanococci Methanocaldococcus
Archaea	Euryarchaeota Methanococci Methanococcus
Archaea	Euryarchaeota Methanococci Methanothermococcus
Archaea	Euryarchaeota Methanococci Methanotorris
Archaea	Euryarchaeota Methanomicrobia Methanocella
Archaea	Euryarchaeota Methanomicrobia Methanococcoides
Archaea	Euryarchaeota Methanomicrobia Methanocorpusculum
Archaea	Euryarchaeota Methanomicrobia Methanoculleus
Archaea	Euryarchaeota Methanomicrobia Methanohalobium
Archaea	Euryarchaeota Methanomicrobia Methanohalophilus
Archaea	Euryarchaeota Methanomicrobia Methanoplanus
Archaea	Euryarchaeota Methanomicrobia Methanoregula
Archaea	Euryarchaeota Methanomicrobia Methanosaeta
Archaea	Euryarchaeota Methanomicrobia Methanosalsum
Archaea	Euryarchaeota Methanomicrobia Methanosarcina
Archaea	Euryarchaeota Methanomicrobia Methanosphaerula
Archaea	Euryarchaeota Methanomicrobia Methanospirillum
Archaea	Euryarchaeota Methanopyri Methanopyrus
Archaea	Euryarchaeota Thermococci Pyrococcus
Archaea	Euryarchaeota Thermococci Thermococcus
Archaea	Euryarchaeota Thermoplasmata Picrophilus
Archaea	Euryarchaeota Thermoplasmata Thermoplasma
Archaea	Korarchaeota Candidatus Korarchaeum
Archaea	Nanoarchaeota Nanoarchaeum
Archaea	Thaumarchaeota Cenarchaeales Cenarchaeum
Archaea	Thaumarchaeota Nitrosopumilales Nitrosopumilus
Bacteria	Acidobacteria Acidobacteriales Acidobacterium
Bacteria	Acidobacteria Acidobacteriales Granulicella
Bacteria	Acidobacteria Acidobacteriales Terriglobus
Bacteria	Acidobacteria Candidatus Chloracidobacterium

Bacteria Acidobacteria Candidatus Koribacter
Bacteria Acidobacteria Solibacteres Candidatus Solibacter
Bacteria Actinobacteria Acidimicrobidae Acidimicrobium
Bacteria Actinobacteria Actinobacteridae Acidothermus
Bacteria Actinobacteria Actinobacteridae Actinoplanes
Bacteria Actinobacteria Actinobacteridae Actinosynnema
Bacteria Actinobacteria Actinobacteridae Amycolatopsis
Bacteria Actinobacteria Actinobacteridae Amycolicicoccus
Bacteria Actinobacteria Actinobacteridae Arcanobacterium
Bacteria Actinobacteria Actinobacteridae Arthrobacter
Bacteria Actinobacteria Actinobacteridae Beutenbergia
Bacteria Actinobacteria Actinobacteridae Bifidobacterium
Bacteria Actinobacteria Actinobacteridae Blastococcus
Bacteria Actinobacteria Actinobacteridae Brachybacterium
Bacteria Actinobacteria Actinobacteridae Catenulispora
Bacteria Actinobacteria Actinobacteridae Cellulomonas
Bacteria Actinobacteria Actinobacteridae Clavibacter
Bacteria Actinobacteria Actinobacteridae Corynebacterium
Bacteria Actinobacteria Actinobacteridae Frankia
Bacteria Actinobacteria Actinobacteridae Gardnerella
Bacteria Actinobacteria Actinobacteridae Geodermatophilus
Bacteria Actinobacteria Actinobacteridae Gordonia
Bacteria Actinobacteria Actinobacteridae Intrasporangium
Bacteria Actinobacteria Actinobacteridae Isoptericola
Bacteria Actinobacteria Actinobacteridae Jonesia
Bacteria Actinobacteria Actinobacteridae Kineococcus
Bacteria Actinobacteria Actinobacteridae Kitasatospora
Bacteria Actinobacteria Actinobacteridae Kocuria
Bacteria Actinobacteria Actinobacteridae Kribbella
Bacteria Actinobacteria Actinobacteridae Kytococcus
Bacteria Actinobacteria Actinobacteridae Leifsonia
Bacteria Actinobacteria Actinobacteridae Microbacterium
Bacteria Actinobacteria Actinobacteridae Micrococcus
Bacteria Actinobacteria Actinobacteridae Microlunatus
Bacteria Actinobacteria Actinobacteridae Micromonospora
Bacteria Actinobacteria Actinobacteridae Mobiluncus
Bacteria Actinobacteria Actinobacteridae Mycobacterium
Bacteria Actinobacteria Actinobacteridae Mycobacterium abscessus
Bacteria Actinobacteria Actinobacteridae Mycobacterium avium complex (MAC)
Bacteria Actinobacteria Actinobacteridae Mycobacterium tuberculosis complex
Bacteria Actinobacteria Actinobacteridae Nakamurella
Bacteria Actinobacteria Actinobacteridae Nocardia
Bacteria Actinobacteria Actinobacteridae Nocardioides
Bacteria Actinobacteria Actinobacteridae Nocardiopsis
Bacteria Actinobacteria Actinobacteridae Propionibacterium
Bacteria Actinobacteria Actinobacteridae Pseudonocardia
Bacteria Actinobacteria Actinobacteridae Renibacterium
Bacteria Actinobacteria Actinobacteridae Rhodococcus
Bacteria Actinobacteria Actinobacteridae Rothia
Bacteria Actinobacteria Actinobacteridae Saccharomonospora
Bacteria Actinobacteria Actinobacteridae Saccharopolyspora
Bacteria Actinobacteria Actinobacteridae Salinispora
Bacteria Actinobacteria Actinobacteridae Sanguibacter
Bacteria Actinobacteria Actinobacteridae Segniliparus
Bacteria Actinobacteria Actinobacteridae Stackebrandtia
Bacteria Actinobacteria Actinobacteridae Streptomyces
Bacteria Actinobacteria Actinobacteridae Streptosporangium
Bacteria Actinobacteria Actinobacteridae Thermobifida
Bacteria Actinobacteria Actinobacteridae Thermobispora
Bacteria Actinobacteria Actinobacteridae Thermomonospora
Bacteria Actinobacteria Actinobacteridae Tropheryma
Bacteria Actinobacteria Actinobacteridae Tsukamurella
Bacteria Actinobacteria Actinobacteridae Verrucosispora
Bacteria Actinobacteria Actinobacteridae Xylanimonas
Bacteria Actinobacteria Coriobacteridae Atopobium
Bacteria Actinobacteria Coriobacteridae Coriobacterium
Bacteria Actinobacteria Coriobacteridae Cryptobacterium
Bacteria Actinobacteria Coriobacteridae Eggerthella
Bacteria Actinobacteria Coriobacteridae Olsenella
Bacteria Actinobacteria Coriobacteridae Slackia
Bacteria Actinobacteria Rubrobacteridae Conexibacter
Bacteria Actinobacteria Rubrobacteridae Rubrobacter
Bacteria Aquificae Aquificales Aquifex
Bacteria Aquificae Aquificales Desulfurobacterium
Bacteria Aquificae Aquificales Hydrogenobacter
Bacteria Aquificae Aquificales Hydrogenobaculum
Bacteria Aquificae Aquificales Persephonella
Bacteria Aquificae Aquificales Sulfurihydrogenibium

Bacteria Aquificae Aquificales Thermocrinis
Bacteria Aquificae Aquificales Thermovibrio
Bacteria Bacteroidetes Bacteroidetes Order II. Incertae sedis Rhodothermus
Bacteria Bacteroidetes Bacteroidetes Order II. Incertae sedis Salinibacter
Bacteria Bacteroidetes Bacteroidia Bacteroides
Bacteria Bacteroidetes Bacteroidia Candidatus Azobacteroides
Bacteria Bacteroidetes Bacteroidia Odoribacter
Bacteria Bacteroidetes Bacteroidia Paludibacter
Bacteria Bacteroidetes Bacteroidia Parabacteroides
Bacteria Bacteroidetes Bacteroidia Porphyromonas
Bacteria Bacteroidetes Bacteroidia Prevotella
Bacteria Bacteroidetes Bacteroidia Tannerella
Bacteria Bacteroidetes Candidatus Amoebophilus
Bacteria Bacteroidetes Cytophagia Cyclobacterium
Bacteria Bacteroidetes Cytophagia Cytophaga
Bacteria Bacteroidetes Cytophagia Dyadobacter
Bacteria Bacteroidetes Cytophagia Leadbetterella
Bacteria Bacteroidetes Cytophagia Marivirga
Bacteria Bacteroidetes Cytophagia Runella
Bacteria Bacteroidetes Cytophagia Spirosoma
Bacteria Bacteroidetes Flavobacteriia Blattabacterium
Bacteria Bacteroidetes Flavobacteriia Candidatus Sulcia
Bacteria Bacteroidetes Flavobacteriia Capnocytophaga
Bacteria Bacteroidetes Flavobacteriia Cellulophaga
Bacteria Bacteroidetes Flavobacteriia Croceibacter
Bacteria Bacteroidetes Flavobacteriia Flavobacteriaceae
Bacteria Bacteroidetes Flavobacteriia Flavobacterium
Bacteria Bacteroidetes Flavobacteriia Fluviicola
Bacteria Bacteroidetes Flavobacteriia Gramella
Bacteria Bacteroidetes Flavobacteriia Krokinobacter
Bacteria Bacteroidetes Flavobacteriia Lacinutrix
Bacteria Bacteroidetes Flavobacteriia Maribacter
Bacteria Bacteroidetes Flavobacteriia Muricauda
Bacteria Bacteroidetes Flavobacteriia Owenweeksia
Bacteria Bacteroidetes Flavobacteriia Riemerella
Bacteria Bacteroidetes Flavobacteriia Robiginitalea
Bacteria Bacteroidetes Flavobacteriia Weeksella
Bacteria Bacteroidetes Flavobacteriia Zobellia
Bacteria Bacteroidetes Flavobacteriia Zunongwangia
Bacteria Bacteroidetes Sphingobacteriia Chitinophaga
Bacteria Bacteroidetes Sphingobacteriia Haliscomenobacter
Bacteria Bacteroidetes Sphingobacteriia Niastella
Bacteria Bacteroidetes Sphingobacteriia Pedobacter
Bacteria Bacteroidetes Sphingobacteriia Saprospira
Bacteria Bacteroidetes Sphingobacteriia Solitalea
Bacteria Bacteroidetes Sphingobacteriia Sphingobacterium
Bacteria Caldiserica Caldisericia Caldisericum
Bacteria candidate division WWE1 Candidatus Cloacamonas
Bacteria Chlamydiae Chlamydiales Candidatus Protochlamydia
Bacteria Chlamydiae Chlamydiales Chlamydia
Bacteria Chlamydiae Chlamydiales Chlamydophila
Bacteria Chlamydiae Chlamydiales Parachlamydia
Bacteria Chlamydiae Chlamydiales Simkania
Bacteria Chlamydiae Chlamydiales Waddlia
Bacteria Chlorobi Chlorobia Chlorobaculum
Bacteria Chlorobi Chlorobia Chlorobium
Bacteria Chlorobi Chlorobia Chloroherpeton
Bacteria Chlorobi Chlorobia Pelodictyon
Bacteria Chlorobi Chlorobia Prosthecochloris
Bacteria Chloroflexi Anaerolineae Anaerolinea
Bacteria Chloroflexi Caldilineae Caldilinea
Bacteria Chloroflexi Chloroflexales Chloroflexus
Bacteria Chloroflexi Chloroflexales Roseiflexus
Bacteria Chloroflexi Dehalococcoidetes Dehalococcoides
Bacteria Chloroflexi Dehalococcoidetes Dehalogenimonas
Bacteria Chloroflexi Herpetosiphonales Herpetosiphon
Bacteria Chloroflexi Sphaerobacteridae Sphaerobacter
Bacteria Chloroflexi Thermomicrobiales Thermomicrobium
Bacteria Chrysiogenetes Chrysiogenales Desulfurispirillum
Bacteria Cyanobacteria Chroococcales
Bacteria Cyanobacteria Chroococcales Acaryochloris
Bacteria Cyanobacteria Chroococcales Cyanothece
Bacteria Cyanobacteria Chroococcales Microcystis
Bacteria Cyanobacteria Chroococcales Synechococcus
Bacteria Cyanobacteria Chroococcales Synechocystis
Bacteria Cyanobacteria Chroococcales Thermosynechococcus
Bacteria Cyanobacteria Gloeobacteria Gloeobacter
Bacteria Cyanobacteria Nostocales Anabaena

Bacteria Cyanobacteria Nostocales Nostoc
Bacteria Cyanobacteria Nostocales Trichormus
Bacteria Cyanobacteria Oscillatoriales Trichodesmium
Bacteria Cyanobacteria Prochlorophytes Prochlorococcus
Bacteria Deferribacteres Deferribacterales Deferribacter
Bacteria Deferribacteres Deferribacterales Deferribacteraceae
Bacteria Deferribacteres Deferribacterales Denitrovibrio
Bacteria Deferribacteres Deferribacterales Flexistipes
Bacteria Deinococcus-Thermus Deinococci Deinococcus
Bacteria Deinococcus-Thermus Deinococci Marinithermus
Bacteria Deinococcus-Thermus Deinococci Meiothermus
Bacteria Deinococcus-Thermus Deinococci Oceanithermus
Bacteria Deinococcus-Thermus Deinococci Thermus
Bacteria Deinococcus-Thermus Deinococci Truepera
Bacteria Dictyoglomi Dictyoglomales Dictyoglomus
Bacteria Elusimicrobia Elusimicrobia Elusimicrobium
Bacteria Elusimicrobia environmental samples
Bacteria Fibrobacteres Fibrobacterales Fibrobacter
Bacteria Firmicutes Bacillales Alicyclobacillus
Bacteria Firmicutes Bacillales Anoxybacillus
Bacteria Firmicutes Bacillales Bacillus
Bacteria Firmicutes Bacillales Bacillus cereus group
Bacteria Firmicutes Bacillales Brevibacillus
Bacteria Firmicutes Bacillales Exiguobacterium
Bacteria Firmicutes Bacillales Geobacillus
Bacteria Firmicutes Bacillales Halobacillus
Bacteria Firmicutes Bacillales Kyrpidia
Bacteria Firmicutes Bacillales Listeria
Bacteria Firmicutes Bacillales Lysinibacillus
Bacteria Firmicutes Bacillales Macrococcus
Bacteria Firmicutes Bacillales Oceanobacillus
Bacteria Firmicutes Bacillales Paenibacillus
Bacteria Firmicutes Bacillales Solibacillus
Bacteria Firmicutes Bacillales Staphylococcus
Bacteria Firmicutes Clostridia Acetobacterium
Bacteria Firmicutes Clostridia Acetohalobium
Bacteria Firmicutes Clostridia Alkaliphilus
Bacteria Firmicutes Clostridia Ammonifex
Bacteria Firmicutes Clostridia Anaerococcus
Bacteria Firmicutes Clostridia Butyrivibrio
Bacteria Firmicutes Clostridia Caldanaerobacter
Bacteria Firmicutes Clostridia Caldicellulosiruptor
Bacteria Firmicutes Clostridia Candidatus Arthromitus
Bacteria Firmicutes Clostridia Candidatus Desulforudis
Bacteria Firmicutes Clostridia Carboxydothermus
Bacteria Firmicutes Clostridia Cellulosilyticum
Bacteria Firmicutes Clostridia Clostridiales
Bacteria Firmicutes Clostridia Clostridium
Bacteria Firmicutes Clostridia Coprothermobacter
Bacteria Firmicutes Clostridia Desulfitobacterium
Bacteria Firmicutes Clostridia Desulfosporosinus
Bacteria Firmicutes Clostridia Desulfotomaculum
Bacteria Firmicutes Clostridia Ethanoligenens
Bacteria Firmicutes Clostridia Eubacterium
Bacteria Firmicutes Clostridia Filifactor
Bacteria Firmicutes Clostridia Finegoldia
Bacteria Firmicutes Clostridia Halanaerobium
Bacteria Firmicutes Clostridia Halothermothrix
Bacteria Firmicutes Clostridia Heliobacterium
Bacteria Firmicutes Clostridia Mahella
Bacteria Firmicutes Clostridia Moorella
Bacteria Firmicutes Clostridia Natranaerobius
Bacteria Firmicutes Clostridia Oscillibacter
Bacteria Firmicutes Clostridia Pelotomaculum
Bacteria Firmicutes Clostridia Peptostreptococcaceae
Bacteria Firmicutes Clostridia Roseburia
Bacteria Firmicutes Clostridia Ruminococcus
Bacteria Firmicutes Clostridia Sulfobacillus
Bacteria Firmicutes Clostridia Symbiobacterium
Bacteria Firmicutes Clostridia Syntrophobotulus
Bacteria Firmicutes Clostridia Syntrophomonas
Bacteria Firmicutes Clostridia Syntrophothermus
Bacteria Firmicutes Clostridia Tepidanaerobacter
Bacteria Firmicutes Clostridia Thermaerobacter
Bacteria Firmicutes Clostridia Thermincola
Bacteria Firmicutes Clostridia Thermoanaerobacter
Bacteria Firmicutes Clostridia Thermoanaerobacterium
Bacteria Firmicutes Clostridia Thermosediminibacter

Bacteria Firmicutes Erysipelotrichi Erysipelothrix
Bacteria Firmicutes Lactobacillales Aerococcus
Bacteria Firmicutes Lactobacillales Carnobacterium
Bacteria Firmicutes Lactobacillales Enterococcus
Bacteria Firmicutes Lactobacillales Lactobacillus
Bacteria Firmicutes Lactobacillales Lactococcus
Bacteria Firmicutes Lactobacillales Leuconostoc
Bacteria Firmicutes Lactobacillales Melissococcus
Bacteria Firmicutes Lactobacillales Oenococcus
Bacteria Firmicutes Lactobacillales Pediococcus
Bacteria Firmicutes Lactobacillales Streptococcus
Bacteria Firmicutes Lactobacillales Tetragenococcus
Bacteria Firmicutes Lactobacillales Weissella
Bacteria Firmicutes Negativicutes Acidaminococcus
Bacteria Firmicutes Negativicutes Selenomonas
Bacteria Firmicutes Negativicutes Veillonella
Bacteria Fusobacteria Fusobacterales Fusobacterium
Bacteria Fusobacteria Fusobacterales Ilyobacter
Bacteria Fusobacteria Fusobacterales Leptotrichia
Bacteria Fusobacteria Fusobacterales Sebaldella
Bacteria Fusobacteria Fusobacterales Streptobacillus
Bacteria Gemmatimonadetes Gemmatimonadales Gemmatimonas
Bacteria Ignavibacteria Ignavibacteria Ignavibacterium
Bacteria Nitrospirae Nitrospirales Leptospirillum
Bacteria Nitrospirae Nitrospirales Thermodesulfovibrio
Bacteria Planctomycetes Phycisphaerae Phycisphaera
Bacteria Planctomycetes Planctomycetia Isosphaera
Bacteria Planctomycetes Planctomycetia Pirellula
Bacteria Planctomycetes Planctomycetia Planctomyces
Bacteria Planctomycetes Planctomycetia Rhodopirellula
Bacteria Proteobacteria Alphaproteobacteria Acetobacter
Bacteria Proteobacteria Alphaproteobacteria Acidiphilium
Bacteria Proteobacteria Alphaproteobacteria Agrobacterium
Bacteria Proteobacteria Alphaproteobacteria Agrobacterium tumefaciens complex
Bacteria Proteobacteria Alphaproteobacteria Anaplasma
Bacteria Proteobacteria Alphaproteobacteria Asticcacaulis
Bacteria Proteobacteria Alphaproteobacteria Azorhizobium
Bacteria Proteobacteria Alphaproteobacteria Azospirillum
Bacteria Proteobacteria Alphaproteobacteria Bartonella
Bacteria Proteobacteria Alphaproteobacteria Beijerinckia
Bacteria Proteobacteria Alphaproteobacteria belli group
Bacteria Proteobacteria Alphaproteobacteria Bradyrhizobium
Bacteria Proteobacteria Alphaproteobacteria Brevundimonas
Bacteria Proteobacteria Alphaproteobacteria Brucella
Bacteria Proteobacteria Alphaproteobacteria Candidatus Hodgkinia
Bacteria Proteobacteria Alphaproteobacteria Candidatus Liberibacter
Bacteria Proteobacteria Alphaproteobacteria Candidatus Midichloria
Bacteria Proteobacteria Alphaproteobacteria Candidatus Pelagibacter
Bacteria Proteobacteria Alphaproteobacteria Candidatus Puniceispirillum
Bacteria Proteobacteria Alphaproteobacteria Caulobacter
Bacteria Proteobacteria Alphaproteobacteria Chelativorans
Bacteria Proteobacteria Alphaproteobacteria Dinoroseobacter
Bacteria Proteobacteria Alphaproteobacteria Ehrlichia
Bacteria Proteobacteria Alphaproteobacteria Erythrobacter
Bacteria Proteobacteria Alphaproteobacteria Gluconacetobacter
Bacteria Proteobacteria Alphaproteobacteria Gluconobacter
Bacteria Proteobacteria Alphaproteobacteria Granulibacter
Bacteria Proteobacteria Alphaproteobacteria Hirschia
Bacteria Proteobacteria Alphaproteobacteria Hyphomicrobium
Bacteria Proteobacteria Alphaproteobacteria Hyphomonas
Bacteria Proteobacteria Alphaproteobacteria Jannaschia
Bacteria Proteobacteria Alphaproteobacteria Ketogulonicigenium
Bacteria Proteobacteria Alphaproteobacteria Magnetococcus
Bacteria Proteobacteria Alphaproteobacteria Magnetospirillum
Bacteria Proteobacteria Alphaproteobacteria Maricaulis
Bacteria Proteobacteria Alphaproteobacteria Mesorhizobium
Bacteria Proteobacteria Alphaproteobacteria Methylobacterium
Bacteria Proteobacteria Alphaproteobacteria Methylocella
Bacteria Proteobacteria Alphaproteobacteria Micavibrio
Bacteria Proteobacteria Alphaproteobacteria Neorickettsia
Bacteria Proteobacteria Alphaproteobacteria Nitrobacter
Bacteria Proteobacteria Alphaproteobacteria Novosphingobium
Bacteria Proteobacteria Alphaproteobacteria Ochrobactrum
Bacteria Proteobacteria Alphaproteobacteria Oligotropha
Bacteria Proteobacteria Alphaproteobacteria Orientia
Bacteria Proteobacteria Alphaproteobacteria Paracoccus
Bacteria Proteobacteria Alphaproteobacteria Parvibaculum
Bacteria Proteobacteria Alphaproteobacteria Parvularcula

Bacteria Proteobacteria Alphaproteobacteria Pelagibacterium
Bacteria Proteobacteria Alphaproteobacteria phagocytophilum group
Bacteria Proteobacteria Alphaproteobacteria Phenylobacterium
Bacteria Proteobacteria Alphaproteobacteria Polymorphum
Bacteria Proteobacteria Alphaproteobacteria Pseudovibrio
Bacteria Proteobacteria Alphaproteobacteria Rhizobium
Bacteria Proteobacteria Alphaproteobacteria Rhodobacter
Bacteria Proteobacteria Alphaproteobacteria Rhodomicrobium
Bacteria Proteobacteria Alphaproteobacteria Rhodopseudomonas
Bacteria Proteobacteria Alphaproteobacteria Rhodospirillum
Bacteria Proteobacteria Alphaproteobacteria Roseobacter
Bacteria Proteobacteria Alphaproteobacteria Ruegeria
Bacteria Proteobacteria Alphaproteobacteria Sinorhizobium
Bacteria Proteobacteria Alphaproteobacteria Sphingobium
Bacteria Proteobacteria Alphaproteobacteria Sphingomonas
Bacteria Proteobacteria Alphaproteobacteria Sphingopyxis
Bacteria Proteobacteria Alphaproteobacteria spotted fever group
Bacteria Proteobacteria Alphaproteobacteria Starkeya
Bacteria Proteobacteria Alphaproteobacteria typhus group
Bacteria Proteobacteria Alphaproteobacteria Wolbachia
Bacteria Proteobacteria Alphaproteobacteria Xanthobacter
Bacteria Proteobacteria Alphaproteobacteria Zymomonas
Bacteria Proteobacteria Betaproteobacteria Achromobacter
Bacteria Proteobacteria Betaproteobacteria Acidovorax
Bacteria Proteobacteria Betaproteobacteria Albidiferax
Bacteria Proteobacteria Betaproteobacteria Alicycliphilus
Bacteria Proteobacteria Betaproteobacteria Aromatoleum
Bacteria Proteobacteria Betaproteobacteria Azoarcus
Bacteria Proteobacteria Betaproteobacteria Azospira
Bacteria Proteobacteria Betaproteobacteria Bordetella
Bacteria Proteobacteria Betaproteobacteria Burkholderia
Bacteria Proteobacteria Betaproteobacteria Burkholderia cepacia complex
Bacteria Proteobacteria Betaproteobacteria Candidatus Accumulibacter
Bacteria Proteobacteria Betaproteobacteria Candidatus Tremblaya
Bacteria Proteobacteria Betaproteobacteria Candidatus Zinderia
Bacteria Proteobacteria Betaproteobacteria Chromobacterium
Bacteria Proteobacteria Betaproteobacteria Collimonas
Bacteria Proteobacteria Betaproteobacteria Comamonas
Bacteria Proteobacteria Betaproteobacteria Cupriavidus
Bacteria Proteobacteria Betaproteobacteria Dechloromonas
Bacteria Proteobacteria Betaproteobacteria Delftia
Bacteria Proteobacteria Betaproteobacteria Gallionella
Bacteria Proteobacteria Betaproteobacteria Herbaspirillum
Bacteria Proteobacteria Betaproteobacteria Herminiimonas
Bacteria Proteobacteria Betaproteobacteria Janthinobacterium
Bacteria Proteobacteria Betaproteobacteria Laribacter
Bacteria Proteobacteria Betaproteobacteria Leptothrix
Bacteria Proteobacteria Betaproteobacteria Methylibium
Bacteria Proteobacteria Betaproteobacteria Methylobacillus
Bacteria Proteobacteria Betaproteobacteria Methylotenera
Bacteria Proteobacteria Betaproteobacteria Methylovorus
Bacteria Proteobacteria Betaproteobacteria Neisseria
Bacteria Proteobacteria Betaproteobacteria Nitrosomonas
Bacteria Proteobacteria Betaproteobacteria Nitrosospira
Bacteria Proteobacteria Betaproteobacteria Polaromonas
Bacteria Proteobacteria Betaproteobacteria Polynucleobacter
Bacteria Proteobacteria Betaproteobacteria Pseudogulbenkiania
Bacteria Proteobacteria Betaproteobacteria pseudomallei group
Bacteria Proteobacteria Betaproteobacteria Pusillimonas
Bacteria Proteobacteria Betaproteobacteria Ralstonia
Bacteria Proteobacteria Betaproteobacteria Ramlibacter
Bacteria Proteobacteria Betaproteobacteria Rubrivivax
Bacteria Proteobacteria Betaproteobacteria Sideroxydans
Bacteria Proteobacteria Betaproteobacteria Taylorella
Bacteria Proteobacteria Betaproteobacteria Thauera
Bacteria Proteobacteria Betaproteobacteria Thiobacillus
Bacteria Proteobacteria Betaproteobacteria Thiomonas
Bacteria Proteobacteria Betaproteobacteria Variovorax
Bacteria Proteobacteria Betaproteobacteria Verminephrobacter
Bacteria Proteobacteria Deltaproteobacteria Anaeromyxobacter
Bacteria Proteobacteria Deltaproteobacteria Bacteriovorax
Bacteria Proteobacteria Deltaproteobacteria Bdellovibrio
Bacteria Proteobacteria Deltaproteobacteria Corallococcus
Bacteria Proteobacteria Deltaproteobacteria Desulfarculus
Bacteria Proteobacteria Deltaproteobacteria Desulfatibacillum
Bacteria Proteobacteria Deltaproteobacteria Desulfobacca
Bacteria Proteobacteria Deltaproteobacteria Desulfobacterium
Bacteria Proteobacteria Deltaproteobacteria Desulfobulbus

Bacteria Proteobacteria Deltaproteobacteria Desulfococcus
Bacteria Proteobacteria Deltaproteobacteria Desulfohalobium
Bacteria Proteobacteria Deltaproteobacteria Desulfomicrobium
Bacteria Proteobacteria Deltaproteobacteria Desulfotalea
Bacteria Proteobacteria Deltaproteobacteria Desulfovibrio
Bacteria Proteobacteria Deltaproteobacteria Desulfurivibrio
Bacteria Proteobacteria Deltaproteobacteria Geobacter
Bacteria Proteobacteria Deltaproteobacteria Haliangium
Bacteria Proteobacteria Deltaproteobacteria Hippea
Bacteria Proteobacteria Deltaproteobacteria Lawsonia
Bacteria Proteobacteria Deltaproteobacteria Myxococcus
Bacteria Proteobacteria Deltaproteobacteria Pelobacter
Bacteria Proteobacteria Deltaproteobacteria Sorangium
Bacteria Proteobacteria Deltaproteobacteria Stigmatella
Bacteria Proteobacteria Deltaproteobacteria Syntrophobacter
Bacteria Proteobacteria Deltaproteobacteria Syntrophus
Bacteria Proteobacteria Epsilonproteobacteria Arcobacter
Bacteria Proteobacteria Epsilonproteobacteria Campylobacter
Bacteria Proteobacteria Epsilonproteobacteria Helicobacter
Bacteria Proteobacteria Epsilonproteobacteria Nautilia
Bacteria Proteobacteria Epsilonproteobacteria Nitratifractor
Bacteria Proteobacteria Epsilonproteobacteria Nitratiruptor
Bacteria Proteobacteria Epsilonproteobacteria Sulfuricurvum
Bacteria Proteobacteria Epsilonproteobacteria Sulfurimonas
Bacteria Proteobacteria Epsilonproteobacteria Sulfurospirillum
Bacteria Proteobacteria Epsilonproteobacteria Sulfurovum
Bacteria Proteobacteria Epsilonproteobacteria Wolinella
Bacteria Proteobacteria Gammaproteobacteria Acidithiobacillus
Bacteria Proteobacteria Gammaproteobacteria Acinetobacter
Bacteria Proteobacteria Gammaproteobacteria Acinetobacter calcoaceticus or baumannii complex
Bacteria Proteobacteria Gammaproteobacteria Actinobacillus
Bacteria Proteobacteria Gammaproteobacteria Aeromonas
Bacteria Proteobacteria Gammaproteobacteria Aggregatibacter
Bacteria Proteobacteria Gammaproteobacteria Alcanivorax
Bacteria Proteobacteria Gammaproteobacteria Aliivibrio
Bacteria Proteobacteria Gammaproteobacteria Alkalilimnicola
Bacteria Proteobacteria Gammaproteobacteria Allochromatium
Bacteria Proteobacteria Gammaproteobacteria Alteromonas
Bacteria Proteobacteria Gammaproteobacteria Azotobacter
Bacteria Proteobacteria Gammaproteobacteria Basfia
Bacteria Proteobacteria Gammaproteobacteria Buchnera
Bacteria Proteobacteria Gammaproteobacteria Candidatus Baumannia
Bacteria Proteobacteria Gammaproteobacteria Candidatus Blochmannia
Bacteria Proteobacteria Gammaproteobacteria Candidatus Carsonella
Bacteria Proteobacteria Gammaproteobacteria Candidatus Hamiltonella
Bacteria Proteobacteria Gammaproteobacteria Candidatus Moranella
Bacteria Proteobacteria Gammaproteobacteria Candidatus Riesia
Bacteria Proteobacteria Gammaproteobacteria Cellvibrio
Bacteria Proteobacteria Gammaproteobacteria Chromohalobacter
Bacteria Proteobacteria Gammaproteobacteria Citrobacter
Bacteria Proteobacteria Gammaproteobacteria Colwellia
Bacteria Proteobacteria Gammaproteobacteria Coxiella
Bacteria Proteobacteria Gammaproteobacteria Cronobacter
Bacteria Proteobacteria Gammaproteobacteria Dichelobacter
Bacteria Proteobacteria Gammaproteobacteria Dickeya
Bacteria Proteobacteria Gammaproteobacteria Edwardsiella
Bacteria Proteobacteria Gammaproteobacteria Enterobacter
Bacteria Proteobacteria Gammaproteobacteria Enterobacter cloacae complex
Bacteria Proteobacteria Gammaproteobacteria Erwinia
Bacteria Proteobacteria Gammaproteobacteria Escherichia
Bacteria Proteobacteria Gammaproteobacteria Ferrimonas
Bacteria Proteobacteria Gammaproteobacteria Francisella
Bacteria Proteobacteria Gammaproteobacteria Frateuria
Bacteria Proteobacteria Gammaproteobacteria Gallibacterium
Bacteria Proteobacteria Gammaproteobacteria Glaciecola
Bacteria Proteobacteria Gammaproteobacteria Haemophilus
Bacteria Proteobacteria Gammaproteobacteria Hahella
Bacteria Proteobacteria Gammaproteobacteria Halomonas
Bacteria Proteobacteria Gammaproteobacteria Halorhodospira
Bacteria Proteobacteria Gammaproteobacteria Halothiobacillus
Bacteria Proteobacteria Gammaproteobacteria Histophilus
Bacteria Proteobacteria Gammaproteobacteria Idiomarina
Bacteria Proteobacteria Gammaproteobacteria Kangiella
Bacteria Proteobacteria Gammaproteobacteria Klebsiella
Bacteria Proteobacteria Gammaproteobacteria Legionella
Bacteria Proteobacteria Gammaproteobacteria Listonella
Bacteria Proteobacteria Gammaproteobacteria Marinobacter
Bacteria Proteobacteria Gammaproteobacteria Marinomonas

Bacteria Proteobacteria Gammaproteobacteria Methylococcus
Bacteria Proteobacteria Gammaproteobacteria Methylomicrobium
Bacteria Proteobacteria Gammaproteobacteria Methylomonas
Bacteria Proteobacteria Gammaproteobacteria Moraxella
Bacteria Proteobacteria Gammaproteobacteria Nitrosococcus
Bacteria Proteobacteria Gammaproteobacteria Oceanimonas
Bacteria Proteobacteria Gammaproteobacteria Pantoea
Bacteria Proteobacteria Gammaproteobacteria Pasteurella
Bacteria Proteobacteria Gammaproteobacteria Pectobacterium
Bacteria Proteobacteria Gammaproteobacteria Photobacterium
Bacteria Proteobacteria Gammaproteobacteria Photorhabdus
Bacteria Proteobacteria Gammaproteobacteria Proteus
Bacteria Proteobacteria Gammaproteobacteria Providencia
Bacteria Proteobacteria Gammaproteobacteria Pseudoalteromonas
Bacteria Proteobacteria Gammaproteobacteria Pseudomonas
Bacteria Proteobacteria Gammaproteobacteria Pseudomonas syringae
Bacteria Proteobacteria Gammaproteobacteria Pseudoxanthomonas
Bacteria Proteobacteria Gammaproteobacteria Psychrobacter
Bacteria Proteobacteria Gammaproteobacteria Psychromonas
Bacteria Proteobacteria Gammaproteobacteria Rahnella
Bacteria Proteobacteria Gammaproteobacteria Saccharophagus
Bacteria Proteobacteria Gammaproteobacteria Salmonella
Bacteria Proteobacteria Gammaproteobacteria Salmonella dublin
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Agona
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Choleraesuis
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Gallinarum or pullorum
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Heidelberg
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Paratyphi B
Bacteria Proteobacteria Gammaproteobacteria Salmonella enterica subsp. enterica serovar Paratyphi C
Bacteria Proteobacteria Gammaproteobacteria Salmonella enteritidis
Bacteria Proteobacteria Gammaproteobacteria Salmonella gallinarum
Bacteria Proteobacteria Gammaproteobacteria Salmonella newport
Bacteria Proteobacteria Gammaproteobacteria Salmonella typhimurium
Bacteria Proteobacteria Gammaproteobacteria Serratia
Bacteria Proteobacteria Gammaproteobacteria Shewanella
Bacteria Proteobacteria Gammaproteobacteria Shigella
Bacteria Proteobacteria Gammaproteobacteria Sodalis
Bacteria Proteobacteria Gammaproteobacteria Stenotrophomonas maltophilia group
Bacteria Proteobacteria Gammaproteobacteria sulfur-oxidizing symbionts
Bacteria Proteobacteria Gammaproteobacteria Teredinibacter
Bacteria Proteobacteria Gammaproteobacteria Thioalkalimicrobium
Bacteria Proteobacteria Gammaproteobacteria Thioalkalivibrio
Bacteria Proteobacteria Gammaproteobacteria Thiomicrospira
Bacteria Proteobacteria Gammaproteobacteria Tolumonas
Bacteria Proteobacteria Gammaproteobacteria Vibrio
Bacteria Proteobacteria Gammaproteobacteria Wigglesworthia
Bacteria Proteobacteria Gammaproteobacteria Xanthomonas
Bacteria Proteobacteria Gammaproteobacteria Xenorhabdus
Bacteria Proteobacteria Gammaproteobacteria Xylella
Bacteria Proteobacteria Gammaproteobacteria Yersinia
Bacteria Spirochaetes Spirochaetales Borrelia
Bacteria Spirochaetes Spirochaetales Borrelia burgdorferi group
Bacteria Spirochaetes Spirochaetales Brachyspira
Bacteria Spirochaetes Spirochaetales Leptospira
Bacteria Spirochaetes Spirochaetales Sphaerochaeta
Bacteria Spirochaetes Spirochaetales Spirochaeta
Bacteria Spirochaetes Spirochaetales Treponema
Bacteria Synergistetes Synergistia Aminobacterium
Bacteria Synergistetes Synergistia Thermanaerovibrio
Bacteria Synergistetes Synergistia Thermovirga
Bacteria Tenericutes Mollicutes 16SrX (Apple proliferation group)
Bacteria Tenericutes Mollicutes 16SrXII (Stolbur group)
Bacteria Tenericutes Mollicutes Acholeplasma
Bacteria Tenericutes Mollicutes Candidatus Phytoplasma asteris
Bacteria Tenericutes Mollicutes Mesoplasma
Bacteria Tenericutes Mollicutes Mycoplasma
Bacteria Tenericutes Mollicutes Ureaplasma
Bacteria Thermobaculum
Bacteria Thermodesulfobacteria Thermodesulfobacteriales Thermodesulfatator
Bacteria Thermodesulfobacteria Thermodesulfobacteriales Thermodesulfobacterium
Bacteria Thermotogae Thermotogales Fervidobacterium
Bacteria Thermotogae Thermotogales Kosmotoga
Bacteria Thermotogae Thermotogales Marinitoga
Bacteria Thermotogae Thermotogales Petrotoga
Bacteria Thermotogae Thermotogales Thermosipho
Bacteria Thermotogae Thermotogales Thermotoga
Bacteria Verrucomicrobia Opitutae Coraliomargarita
Bacteria Verrucomicrobia Opitutae Opitutus

Bacteria Verrucomicrobia unclassified Verrucomicrobia Methylacidiphilum
Bacteria Verrucomicrobia Verrucomicrobiae Akkermansia

Eukaryota Alveolata Apicomplexa Babesia
Eukaryota Alveolata Apicomplexa Cryptosporidium
Eukaryota Alveolata Apicomplexa Neospora
Eukaryota Alveolata Apicomplexa Plasmodium (Laverania)
Eukaryota Alveolata Apicomplexa Plasmodium (Plasmodium)
Eukaryota Alveolata Apicomplexa Plasmodium (Vinckeia)
Eukaryota Alveolata Apicomplexa Theileria
Eukaryota Alveolata Apicomplexa Toxoplasma
Eukaryota Alveolata Ciliophora Ichthyophthirius
Eukaryota Alveolata Ciliophora Paramecium
Eukaryota Alveolata Ciliophora Tetrahymena
Eukaryota Alveolata Perkinsea Perkinsus
Eukaryota Amoebozoa Archamoebae Entamoeba
Eukaryota Amoebozoa Mycetozoa Dictyostelium
Eukaryota Amoebozoa Mycetozoa Polysphondylium
Eukaryota Choanoflagellida Codonosigidae Monosiga
Eukaryota Choanoflagellida Salpingoecidae Salpingoeca
Eukaryota Diplomonadida Hexamitidae Giardia
Eukaryota Euglenozoa Kinetoplastida Duttonella
Eukaryota Euglenozoa Kinetoplastida Leishmania
Eukaryota Euglenozoa Kinetoplastida Leishmania braziliensis species complex
Eukaryota Euglenozoa Kinetoplastida Nannomonas
Eukaryota Euglenozoa Kinetoplastida Schizotrypanum
Eukaryota Euglenozoa Kinetoplastida Trypanosoma
Eukaryota Fungi Chytridiomycota Batrachochytrium
Eukaryota Fungi Dikarya Ajellomyces
Eukaryota Fungi Dikarya Arthroderma
Eukaryota Fungi Dikarya Aspergillus
Eukaryota Fungi Dikarya Botryotinia
Eukaryota Fungi Dikarya Candida
Eukaryota Fungi Dikarya Chaetomium
Eukaryota Fungi Dikarya Clavispora
Eukaryota Fungi Dikarya Coccidioides
Eukaryota Fungi Dikarya Colletotrichum
Eukaryota Fungi Dikarya Coprinopsis
Eukaryota Fungi Dikarya Cordyceps
Eukaryota Fungi Dikarya Debaryomyces
Eukaryota Fungi Dikarya Emericella
Eukaryota Fungi Dikarya Eremothecium
Eukaryota Fungi Dikarya Exophiala
Eukaryota Fungi Dikarya Filobasidiella or Cryptococcus neoformans species complex
Eukaryota Fungi Dikarya Fusarium oxysporum species complex
Eukaryota Fungi Dikarya Gibberella
Eukaryota Fungi Dikarya Glarea
Eukaryota Fungi Dikarya Glomerella
Eukaryota Fungi Dikarya Grosmannia
Eukaryota Fungi Dikarya Hypocrea
Eukaryota Fungi Dikarya Kazachstania
Eukaryota Fungi Dikarya Kluyveromyces
Eukaryota Fungi Dikarya Komagataella
Eukaryota Fungi Dikarya Laccaria
Eukaryota Fungi Dikarya Lachancea
Eukaryota Fungi Dikarya Leptosphaeria maculans complex
Eukaryota Fungi Dikarya Lodderomyces
Eukaryota Fungi Dikarya Magnaporthe
Eukaryota Fungi Dikarya Malassezia
Eukaryota Fungi Dikarya Melampsora
Eukaryota Fungi Dikarya Metarhizium
Eukaryota Fungi Dikarya Meyerozyma
Eukaryota Fungi Dikarya Millerozyma
Eukaryota Fungi Dikarya mitosporic Nakaseomyces
Eukaryota Fungi Dikarya Mixia
Eukaryota Fungi Dikarya Moniliophthora
Eukaryota Fungi Dikarya Myceliophthora
Eukaryota Fungi Dikarya Naumovozyma
Eukaryota Fungi Dikarya Nectria haematococca complex
Eukaryota Fungi Dikarya Neosartorya
Eukaryota Fungi Dikarya Neurospora
Eukaryota Fungi Dikarya Ogataea
Eukaryota Fungi Dikarya Orbilia
Eukaryota Fungi Dikarya Paracoccidioides
Eukaryota Fungi Dikarya Penicillium
Eukaryota Fungi Dikarya Penicillium chrysogenum complex
Eukaryota Fungi Dikarya Phaeosphaeria
Eukaryota Fungi Dikarya Piriformospora

Eukaryota Fungi Dikarya Podospora
Eukaryota Fungi Dikarya Postia
Eukaryota Fungi Dikarya Puccinia
Eukaryota Fungi Dikarya Pyrenophora
Eukaryota Fungi Dikarya Rhodotorula
Eukaryota Fungi Dikarya Saccharomyces
Eukaryota Fungi Dikarya Scheffersomyces
Eukaryota Fungi Dikarya Schizophyllum
Eukaryota Fungi Dikarya Schizosaccharomyces
Eukaryota Fungi Dikarya Sclerotinia
Eukaryota Fungi Dikarya Serpula
Eukaryota Fungi Dikarya Sordaria
Eukaryota Fungi Dikarya Spathaspora
Eukaryota Fungi Dikarya Sporisorium
Eukaryota Fungi Dikarya Talaromyces
Eukaryota Fungi Dikarya Tetrapisispora
Eukaryota Fungi Dikarya Thielavia
Eukaryota Fungi Dikarya Torulaspora
Eukaryota Fungi Dikarya Trichoderma
Eukaryota Fungi Dikarya Trichophyton
Eukaryota Fungi Dikarya Tuber
Eukaryota Fungi Dikarya Uncinocarpus
Eukaryota Fungi Dikarya Ustilago
Eukaryota Fungi Dikarya Vanderwaltozyma
Eukaryota Fungi Dikarya Verticillium
Eukaryota Fungi Dikarya Yarrowia
Eukaryota Fungi Dikarya Zygosaccharomyces
Eukaryota Fungi Dikarya Zymoseptoria
Eukaryota Fungi Fungi incertae sedis Rhizopus
Eukaryota Fungi Microsporidia Encephalitozoon
Eukaryota Fungi Microsporidia Enterocytozoon
Eukaryota Fungi Microsporidia Nematocida
Eukaryota Fungi Microsporidia Nosema
Eukaryota Heterolobosea Schizopyrenida Naegleria
Eukaryota Ichthyosporea Capsaspora
Eukaryota Metazoa Arthropoda Acromyrmex
Eukaryota Metazoa Arthropoda Anopheles
Eukaryota Metazoa Arthropoda Apis
Eukaryota Metazoa Arthropoda Atta
Eukaryota Metazoa Arthropoda Bombyx
Eukaryota Metazoa Arthropoda Camponotus
Eukaryota Metazoa Arthropoda Culex
Eukaryota Metazoa Arthropoda Danaus
Eukaryota Metazoa Arthropoda Daphnia
Eukaryota Metazoa Arthropoda Drosophila
Eukaryota Metazoa Arthropoda Harpegnathos
Eukaryota Metazoa Arthropoda Hawaiian Drosophila
Eukaryota Metazoa Arthropoda Ixodes
Eukaryota Metazoa Arthropoda Pediculus
Eukaryota Metazoa Arthropoda Solenopsis
Eukaryota Metazoa Arthropoda Sophophora
Eukaryota Metazoa Arthropoda Stegomyia
Eukaryota Metazoa Arthropoda Tribolium
Eukaryota Metazoa Chordata Ailuropoda
Eukaryota Metazoa Chordata Anolis
Eukaryota Metazoa Chordata Bos
Eukaryota Metazoa Chordata Branchiostoma
Eukaryota Metazoa Chordata Callithrix
Eukaryota Metazoa Chordata Canis
Eukaryota Metazoa Chordata Cavia
Eukaryota Metazoa Chordata Ciona
Eukaryota Metazoa Chordata Cricetulus
Eukaryota Metazoa Chordata Danio
Eukaryota Metazoa Chordata Equus
Eukaryota Metazoa Chordata Gallus
Eukaryota Metazoa Chordata Gasterosteus
Eukaryota Metazoa Chordata Gorilla
Eukaryota Metazoa Chordata Heterocephalus
Eukaryota Metazoa Chordata Homo
Eukaryota Metazoa Chordata Ictidomys
Eukaryota Metazoa Chordata Latimeria
Eukaryota Metazoa Chordata Loxodonta
Eukaryota Metazoa Chordata Macaca
Eukaryota Metazoa Chordata Meleagris
Eukaryota Metazoa Chordata Monodelphis
Eukaryota Metazoa Chordata Mus
Eukaryota Metazoa Chordata Myotis
Eukaryota Metazoa Chordata Nomascus

Eukaryota Metazoa Chordata Oikopleura
Eukaryota Metazoa Chordata Oreochromis
Eukaryota Metazoa Chordata Ornithorhynchus
Eukaryota Metazoa Chordata Oryctolagus
Eukaryota Metazoa Chordata Oryzias
Eukaryota Metazoa Chordata Otolemur
Eukaryota Metazoa Chordata Pan
Eukaryota Metazoa Chordata Pongo
Eukaryota Metazoa Chordata Rattus
Eukaryota Metazoa Chordata Sarcophilus
Eukaryota Metazoa Chordata Silurana
Eukaryota Metazoa Chordata Sus
Eukaryota Metazoa Chordata Taeniopygia
Eukaryota Metazoa Chordata Takifugu
Eukaryota Metazoa Chordata Tetraodon
Eukaryota Metazoa Cnidaria Nematostella
Eukaryota Metazoa Echinodermata Strongylocentrotus
Eukaryota Metazoa Nematoda Brugia
Eukaryota Metazoa Nematoda Caenorhabditis
Eukaryota Metazoa Nematoda Loa
Eukaryota Metazoa Nematoda Pristionchus
Eukaryota Metazoa Nematoda Trichinella
Eukaryota Metazoa Placozoa Trichoplax
Eukaryota Metazoa Platyhelminthes Clonorchis
Eukaryota Metazoa Platyhelminthes Schistosoma
Eukaryota Metazoa Porifera Amphimedon
Eukaryota Parabasalia Trichomonadida Trichomonas
Eukaryota stramenopiles Bacillariophyta Phaeodactylum
Eukaryota stramenopiles Bacillariophyta Thalassiosira
Eukaryota stramenopiles Blastocystis
Eukaryota stramenopiles Oomycetes Phytophthora
Eukaryota stramenopiles Pelagophyceae Aureococcus
Eukaryota stramenopiles PX clade Ectocarpus
Eukaryota Viridiplantae Chlorophyta Chlamydomonas
Eukaryota Viridiplantae Chlorophyta Chlorella
Eukaryota Viridiplantae Chlorophyta Micromonas
Eukaryota Viridiplantae Chlorophyta Ostreococcus
Eukaryota Viridiplantae Chlorophyta Volvox
Eukaryota Viridiplantae Streptophyta Arabidopsis
Eukaryota Viridiplantae Streptophyta Brachypodium
Eukaryota Viridiplantae Streptophyta Glycine
Eukaryota Viridiplantae Streptophyta Oryza
Eukaryota Viridiplantae Streptophyta Physcomitrella
Eukaryota Viridiplantae Streptophyta Populus
Eukaryota Viridiplantae Streptophyta Ricinus
Eukaryota Viridiplantae Streptophyta Selaginella
Eukaryota Viridiplantae Streptophyta Sorghum
Eukaryota Viridiplantae Streptophyta Vitis