

Current Protocols in Protein Science

Unit 2.5

Sequence Similarity Searching

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ABSTRACT

Sequence similarity searching has become an important part of the daily routine of molecular biologists, bioinformaticians and biophysicists. With the rapidly growing sequence databanks, this computational approach is commonly applied to determine functions and structures of unannotated sequences, to investigate relationships between sequences, and to construct phylogenetic trees. We introduce arguably the most popular BLAST-based family of the sequence similarity search tools. We explain basic concepts related to the sequence alignment and demonstrate how to search the current databanks using website versions of BLASTP, PSI-BLAST and BLASTN. We also describe the standalone BLAST+ tool. Moreover, this unit discusses the inputs, parameter settings and outputs of these tools. Lastly, we cover recent advances in the sequence similarity searching, focusing on the fast MMseqs2 method.

Keywords: sequence similarity searching; alignment; BLAST; BLASTP, BLASTN, PSSM; MMseqs2.

INTRODUCTION TO SEQUENCE SIMILARITY SEARCHING AND BLAST PROGRAMS

Identification of similar sequences in large sequence databanks is typically the first step in analysis of new protein, DNA and RNA sequences. Structural and functional annotations of the similar sequences can be used to infer functions and structural features of the new sequences. Many databanks, such as NCBI's GenBank (Benson et al., 2018), EMBL's Nucleotide Sequence Database (Kulikova et al., 2007), Universal Protein Resource (UniProt) (Boutet et al., 2016; The UniProt, 2017), and Protein Data Bank (PDB) (Berman et al., 2000; Rose et al., 2017), provide a rich source of millions of sequences and annotations. More specifically, GenBank provides access to a comprehensive and non-redundant set of reference sequences, the Reference Sequence Database (RefSeq) (O'Leary et al., 2016). As of January 2018, RefSeq includes over 102 million protein sequences and over 21 million transcripts. UniProt is the main database of protein sequences and functional annotations. As of January 2018, it includes 556 thousand manually reviewed proteins and over 107 million computationally analyzed proteins. PDB provides access to 137,000 experimentally determined protein structures. Moreover, NCBI provides the nr database, a comprehensive collection of non-redundant protein sequences from RefSeq, UniProt, PDB and other protein databases, to facilitate searching for similar sequences.

Many algorithms that can be used to search for similar sequences were developed over the last three decades. They include FASTA (Pearson and Lipman, 1988), ClustalW (Larkin et al., 2007; Thompson et al., 1994), HMMER (Johnson et al., 2010), MMseqs2 (Steinegger and Soding, 2017), and Basic Local Alignment Search Tool (BLAST) (Altschul et al., 1990; Camacho et al., 2009). BLAST is arguably the most popular tool. The article that has introduced this algorithm was cited over 70 thousand times (source: Google Scholar on Feb. 22, 2018). BLAST performs and scores sequence alignment to find similar nucleotide or protein sequences for a given query sequence. The search is executed against a large database (typically the nr database) and the algorithm also quantifies statistical significance of the scored output matches. The family of the BLAST programs offers a variety of options that address searching of nucleotide and protein chains, see Table 1. One of the key factors that contribute to the popularity of BLAST is its omnipresent availability. BLAST algorithms are available in two main flavors: 1) WEB BLAST (Johnson et al., 2008), a web-based application that is accessible using any of the major web browsers for which the computations are performed on the webserver side; and 2) BLAST+ (Camacho et al., 2009), a standalone software that can be installed and run on a local computer. The latter option is particularly attractive for users who would like to include BLAST into a larger computational pipeline. The WEB BLAST can be accessed through the NCBI website at <https://blast.ncbi.nlm.nih.gov/Blast.cgi>. BLAST+ can be downloaded from <ftp://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/LATEST/>. The programs listed in Table 1 can be found in both WEB BLAST and BLAST+. WEB BLAST also provides access to a variety of search databases including the nr, RefSeq, SWISS-PROT, PDB, human genome, mouse genome etc.

Next, we describe the WEB BLAST and BLAST+ for proteins, followed by the BLAST programs for the nucleotide chains. We conclude this Unit with a discussion of recently developed alternatives to BLAST.

Table 1. Summary of the BLAST programs.

Programs	Query sequence	Target database	Description
BLASTP suite	Protein	Protein	To search protein sequences against a protein database. Includes BLASTP, Quick BLASTP, PSI-BLAST, PHI-BLAST and DELTA-BLAST.
TBLASTN	Protein	Translated Nucleotide	To identify nucleotide sequences encoding proteins similar to the query protein.
BLASTN suite	Nucleotide	Nucleotide	To search nucleotide sequences against a nucleotide database. Includes MEGABLAST, discontinuous MEGABLAST and BLASTN.
BLASTX	Translated Nucleotide	Protein	To identify potential protein products encoded by a nucleotide query.
TBLASTX	Translated Nucleotide	Translated Nucleotide	To identify nucleotide sequences similar to the query transcript based on their coding potential.

BLASTP SUITE - BLAST AGAINST PROTEIN DATABASES

The sequence similarity search performed by BLAST has roots in early sequence alignment algorithms (Needleman and Wunsch, 1970; Smith and Waterman, 1981). The main difference is that BLAST performs a heuristic search that is characterized by a much faster convergence to a solution. The search maximizes a score that quantifies similarity between sequences, given a particular scoring matrix and gap penalty. The widely used scoring matrices for the protein sequences include Point Accepted Mutation (PAM) matrices (Schwarz and Dayhoff, 1979) and BLOcks SUBstitution Matrices (BLOSUM) (Altschul, 1991). These matrices quantify similarities between all pairs of amino acids, such that pairs of the same or similar amino acids have high scores while pairs of dissimilar amino acids are associated with low scores. Figure 1 gives one of the most popular BLOSUM62 scoring matrix. This symmetrical matrix has 20 rows and 20 columns. The number in the i^{th} row and j^{th} column is the similarity score for the pair of i^{th} and j^{th} amino acids. The gap penalty is for gaps (openings) that are often inserted into one of the aligned sequences to maximize the similarity score. The gap penalty is larger for a new gap when compared to an extension of an already existing gap. We show examples of alignments with gaps and discuss how to setup the gap penalty values in the “BLASTN Suite” and “Recent Advances” sections. BLAST can be parametrized to use different scoring matrices and gap penalties, resulting in different alignments for the same sequences.

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V	B	Z	X
A	4	-1	-2	-2	0	-1	-1	0	-2	-1	-1	-1	-2	-1	1	0	-3	-2	0	-2	-1	0	-1
R	-1	5	0	-2	-3	1	0	-2	0	-3	-2	2	-1	-3	-2	-1	-1	-3	-2	-3	-1	0	-1
N	-2	0	6	1	-3	0	0	0	1	-3	-3	0	-2	-3	-2	1	0	-4	-2	-3	3	0	-1
D	-2	-2	1	6	-3	0	2	-1	-1	-3	-4	-1	-3	-3	-1	0	-1	-4	-3	-3	4	1	-1
C	0	-3	-3	-3	9	-3	-4	-3	-3	-1	-1	-3	-1	-2	-3	-1	-1	-2	-2	-1	-3	-3	-2
Q	-1	1	0	0	-3	5	2	-2	0	-3	-2	1	0	-3	-1	0	-1	-2	-1	-2	0	3	-1
E	-1	0	0	2	-4	2	5	-2	0	-3	-3	1	-2	-3	-1	0	-1	-3	-2	-2	1	4	-1
G	0	-2	0	-1	-3	-2	-2	6	-2	-4	-4	-2	-3	-3	-2	0	-2	-2	-3	-3	-1	-2	-1
H	-2	0	1	-1	-3	0	0	-2	8	-3	-3	-1	-2	-1	-2	-1	-2	-2	-2	-3	0	0	-1
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4	2	3	1	0	-3	-2	-1	-3	-1	3	-3	-3	-1
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4	-2	2	0	-3	-2	-1	-2	-1	1	-4	-3	-1
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5	-1	-3	-1	0	-1	-3	-2	-2	0	1	-1
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5	0	-2	-1	-1	-1	-1	1	-3	-1	-1
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6	-4	-2	-2	1	3	-1	-3	-3	-1
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7	-1	-1	-4	-3	-2	-2	-1	-2
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4	1	-3	-2	-2	0	0	0
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-2	-1	1	5	-2	-2	0	-1	-1	0	-1
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11	2	-3	-4	-3	-2
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	-1	-3	-2	-1
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4	-3	-2	-1
B	-2	-1	3	4	-3	0	1	-1	0	-3	-4	0	-3	-3	-2	0	-1	-4	-3	-3	4	1	-1
Z	-1	0	0	1	-3	3	4	-2	0	-3	-3	1	-1	-3	-1	0	-1	-3	-2	-2	1	4	-1
X	0	-1	-1	-1	-2	-1	-1	-1	-1	-1	-1	-1	-1	-2	0	0	-2	-1	-1	-1	-1	-1	-1

Figure 1. The BLOSUM62 scoring matrix.

The input/query sequence and the sequences in the various databases are stored in a common format to expedite the search. The most popular sequence format is FASTA. This format was first introduced in the FASTA program for sequence alignment (Lipman and Pearson, 1985) and was adopted as a standard for representing nucleotide and protein sequences. Virtually all sequences databases support this format. FASTA is a text based format for both protein and nucleotide sequences. It uses multiple lines to store a single sequence. The first line starts with “>” symbol which is followed by comments that typically name and provide information about the sequence; multiple comment lines can be used. The subsequent lines give the one-letter encoded protein sequence. An example FASTA formatted sequence for the human arrestin protein follows:

```
>sp|P36575|ARRC_HUMAN Arrestin-C OS=Homo sapiens
MSKVFKKTSNGKLSIYLGKRDFVDHVDTVPEPIDGVVLVDPEYLYLKRKLFVMLTCAFQRYG
RDDLEVIQLTFRKDLVQTLQVPAESSSPQGPLTVLQERLLHKLGDNAYPFTLQMTNL
PCSVTLQPGPEDAGKPCGIDFEVKSFCENPEETVSKRDYVRLVVRKVQFAPPEAGFGPS
AQTIRRFLLSAQPLQLQAWMDREVHYHGEPISVNVNSINNCTNKVIKKIKISVDQITDQVVL
YSLDKYTKTVFIQEFETETVAANSFSQSFAVTPILAASCQKRGGLALDGLKHKHEDTNLASS
TIIRPGMDKELLGILVSYKVRVNLMSVCGGILGDLTASDVGVLPVLIHPKPSHEAASS
EDIVIEEFTRKGEESQKAVEAEGDEGS
```

BLASTP suite searches protein sequences against a protein database. It includes five programs: BLASTP, Quick BLASTP, PSI-BLAST (Position-Specific Iterative BLAST), PHI-BLAST (Pattern-Hit Initiated BLAST) and DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST). BLASTP is a generic purpose protein sequence alignment program that compares a query protein sequence to sequences in a specific protein database. Quick BLASTP is an accelerated (faster) version of BLASTP that is specifically coupled with the nr database. PSI-BLAST (Altschul et al., 1997) finds more distant relatives (a wider range of similar sequences) by performing the BLAST search iteratively. It also produces the position specific scoring matrix (PSSM). We explain further details and how to use PSI-BLAST in the next section. PHI-BLAST (Zhang et al., 1998) performs search that is limited to alignments that match a specific pattern in the query sequence. Finally, DELTA-BLAST (Boratyn et al., 2012)s uses pre-computed PSSMs derived from the Conserved Domain Database (Marchler-Bauer et al., 2011) to provide stronger detection of remote homologs.

The web interface of the BLASTP suite, which is shown in Figure 2, includes three parts:

- (1) “Enter Query Sequence” box facilitates inputting of the query sequence(s). The sequence(s) can be provided either directly using the text box or they can be uploaded from a file. The sequences can be provided either directly in the FASTA format or the query protein can be identified with either accession number or NCBI gi number. The accession number and NCBI gi number can be found in the NCBI databases. User can also opt to align part of the query sequence that is defined with the query subrange. Finally, BLASTP can be used to align two query proteins with each other using the “Align two or more sequences” checkbox.
- (2) “Choose Search Set” box allows for selection of the target databases and organisms. Eight databases are currently available including nr, RefSeq, PDB and SWISS-PROT (Bairoch and Apweiler, 2000). List of organisms is optional and it should be used to limit the results to this selection of species.
- (3) “Program Selection” box offers choice of the five BLASTP suite programs where BLASP is selected by default.

Figure 2 gives a query that concerns human arrestin protein which is set up to be searched against the nr database using the default BLASTP program.

The screenshot displays the BLAST web interface for the BLASTP suite. At the top, there are navigation links: Home, Recent Results, Saved Strategies, and Help. The main heading is "Standard Protein BLAST". Below this, there are tabs for different BLAST programs: blastn, blastp, blastx, tblastn, and tblastx. The "blastp" tab is selected.

The "Enter Query Sequence" section contains a text box with the following FASTA sequence:


```
>sp|P36575|ARRC_HUMAN Arrestin-C OS=Homo sapiens GN=ARR3 PE=1 SV=2
MSKVFKKTSNGKLSIYLGKRDVFDHVDTVPEIDGVVLDPEYLKCRKLFVMLTCAFYRG
RDDLEVI GLTKRDLVYQTLQVVPASSSPQGPLTVLQERLLHKLGDNAYPFTLQMVTNL
PCSVTLQPGPEDAGKPCGIDFEVKSFCENPEETVSKRDYVRLVVRKVQFAPPEAGPGPS
AQTIRRFLLSAQPLQLQAWMDREVHYHGEPISVNVSIINNTNKVKKIKISVDQITDVLV
```

 To the right of the text box is a "Clear" button and a "Query subrange" section with "From" and "To" input fields. Below the text box is an "Or, upload file" section with a "Browse..." button. A "Job Title" field contains the text "sp|P36575|ARRC_HUMAN Arrestin-C OS=Homo sapiens...". Below the job title is a checkbox for "Align two or more sequences".

The "Choose Search Set" section includes a "Database" dropdown menu set to "Non-redundant protein sequences (nr)". Below this are optional fields for "Organism" (with a search box and "Exclude" checkbox), "Exclude" (with checkboxes for "Models (XM/XP)" and "Uncultured/environmental sample sequences"), and "Entrez Query" (with a search box and "Create custom database" link).

The "Program Selection" section shows a list of algorithms with radio buttons:

- Quick BLASTP (Accelerated protein-protein BLAST) [New]
- blastp (protein-protein BLAST)** (selected)
- PSI-BLAST (Position-Specific Iterated BLAST)
- PHI-BLAST (Pattern Hit Initiated BLAST)
- DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)

 Below the list is a "Choose a BLAST algorithm" button.

At the bottom, there is a "BLAST" button and a search summary: "Search database Non-redundant protein sequences (nr) using Blastp (protein-protein BLAST)". There is also a checkbox for "Show results in a new window".

Figure 2. Web interface for the BLASTP suite.

The BLAST parameters can be adjusted using a link at the bottom of the web interface page. This is optional and in most cases BLAST is used with pre-selected default parameters, which are shown in Figure 3. Using the parameters menu users can restrict the maximal number of aligned (similar) target sequences (default value is 100) and can set the maximal E-value. The E-value quantifies statistical significance of the similarity where lower value corresponds to more significant similarity; the default value is 10, which means that about 10 of the similar sequences are expected to be found by chance (Karlin and Altschul, 1990). Users can also influence key parameters that govern the sequence similarity search. BLAST uses a filter-based technique to speed up the search for similar sequences. Instead of aligning the sequences directly it finds word matches between the query and database sequences (filter step) and then it extends each word match to a gapped alignment (alignment step). Use of smaller word sizes makes the search more sensitive. The default word size for proteins is set to 6. The “maximal matches in a query range” parameter limits the number of matches and has the default value of 0, which means that there is no limit. The default scoring matrix is BLOSUM62 (Figure 1) and the default setup of the gap penalties is 11 for opening the gap and 1 for the extension of the gap. Different scoring matrices are designed to detect similarities among sequences that diverge by differing degrees (Altschul, 1991; Altschul, 1993). BLOSUM62 is preferred for the detection of weak similarities (Henikoff and Henikoff, 1992). Other matrices are more suitable for specific scenarios. For instance, BLOSUM45 is designed to detect long and weak alignments (Choudhuri, 2014). The “Filters and Masking” section in Figure 3 allows for masking low complexity regions in the protein sequences when the top option is checked. This means that the low complexity regions will not be used in the alignment. If the second option “Mask for lookup table only” is checked then the low complexity regions will be masked only for constructing the lookup table or words list that are used in the filter step while the alignment step is performed without masking. The third option masks lower case letters in the query FASTA sequence; this allows user to choose which parts of the sequence should not be aligned.

The screenshot displays the 'Algorithm parameters' section of the BLASTP suite interface. It is organized into three main sections:

- General Parameters:**
 - Max target sequences:** A dropdown menu set to 100. Below it is the text: "Select the maximum number of aligned sequences to display".
 - Short queries:** A checked checkbox labeled "Automatically adjust parameters for short input sequences".
 - Expect threshold:** A text input field set to 10.
 - Word size:** A dropdown menu set to 6.
 - Max matches in a query range:** A text input field set to 0.
- Scoring Parameters:**
 - Matrix:** A dropdown menu set to BLOSUM62.
 - Gap Costs:** A dropdown menu showing "Existence: 11 Extension: 1".
 - Compositional adjustments:** A dropdown menu set to "Conditional compositional score matrix adjustment".
- Filters and Masking:**
 - Filter:** An unchecked checkbox labeled "Low complexity regions".
 - Mask:** Two unchecked checkboxes: "Mask for lookup table only" and "Mask lower case letters".

At the top right of the interface, there is a link labeled "Restore default search parameters".

Figure 3. Parameters of the BLASTP suite.

We run the query shown in Figures 2 and 3 and the results are summarized in Figures 4, 5 and 6. The webpage that gives the search results is divided into four parts:

- (1) General information that is located at the top includes query identifier and description, target database name and description as well as a detailed summary of key search parameters. These parameters include word size, E-value, gap penalties, scoring matrix, number of sequences scanned, date and time of the request, and about a dozen of other statistics.
- (2) “Graphic Summary” box provides a simplified, graphical representation of the alignment of the query sequence with the top scoring similar sequences (Figure 4). The summary includes the alignment score and E-value for each aligned sequence. Higher alignment score implies higher degree of similarity between the query sequence and a given hit sequence. The E-value quantifies the number of hits that are expected to happen by chance. The multiple alignment for several top hits for our query protein is given in Figure 5. The conserved regions are shown in red. The multiple alignment can be downloaded using a wide variety of formats including FASTA, clustal, phylip, nexus and ASN.1. The “Graphic Summary” box also provides putative conserved domains that can be used to classify the query protein (see the top part of Figure 4). Users can find further details about these conserved domains by clicking on the image. This opens a new webpage and gives a list of domain hits with their sequences and Pfam or smart accession numbers. We find three hits for the arrestin example, each with 2 domains (Figure 6). The first domain is found in the 14 to 171 sequence segment in the query sequence chain and it corresponds to Pfam id 00339 (Arrestin N-terminal domain). The second domain stretches between positions 193 to 353 and concerns Pfam id 02752 (Arrestin C-terminal domain) or alternatively smart id 01017.
- (3) “Descriptions” box provides additional details for each aligned sequence. These details include it’s accession number, alignment score, E-value, query coverage and identity with the query protein that is expressed in percentage points. The query coverage is the fraction of the query length that is included in the aligned segments. The aligned sequences are arranged by their E-values in the ascending order.
- (4) “Alignments” box provides side-by-side alignments between the query sequence and the top hits.

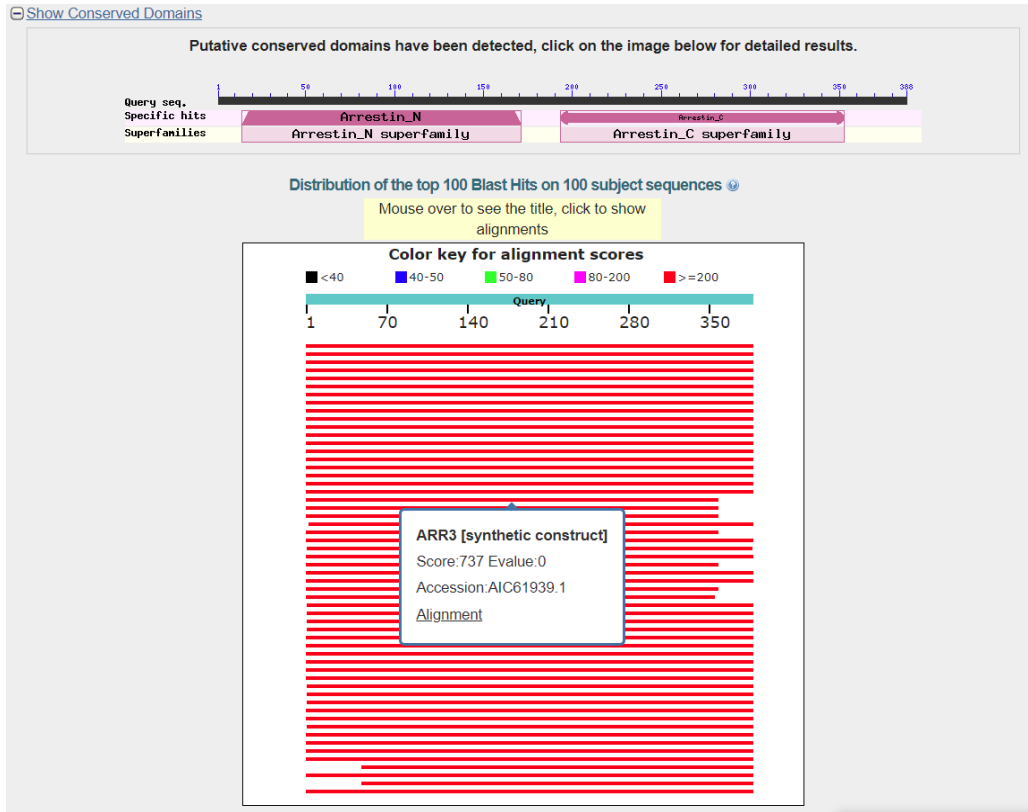


Figure 4. Graphic summary box generated by the BLASTP suite for human arrestin protein.

<input checked="" type="checkbox"/> Query_331358	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> NP_004303	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> AAC78395	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> AAB84302	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GALTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> EAX05349	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> CAG33130	34	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> XP_002834739	34	DGVVLDPEYLNKGRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> XP_003820212	34	DGVVLDPEYLNKGRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> XP_003272725	34	DGVVLDPEYLNKGRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> XP_004064375	34	DGVVLDPEYLNKGRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> PNI41816	34	DGVVLDPEYLNKGRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	113
<input checked="" type="checkbox"/> XP_016885007	49	DGVVLDPEYLNKCRKLFV MLTCAF RYGRDDLEVI GLTFR KDLYVQTLQV VP AESS SPQ GPLTVLQ ER LLHKLGD NAY PFT	128

Figure 5. Multiple alignment generated by the BLASTP suite for the human arrestin proteins. The first column lists the accession identifiers of several top hit sequences. The second and last column are positions of sequences. The third column is the alignment where conserved regions are shown in red.

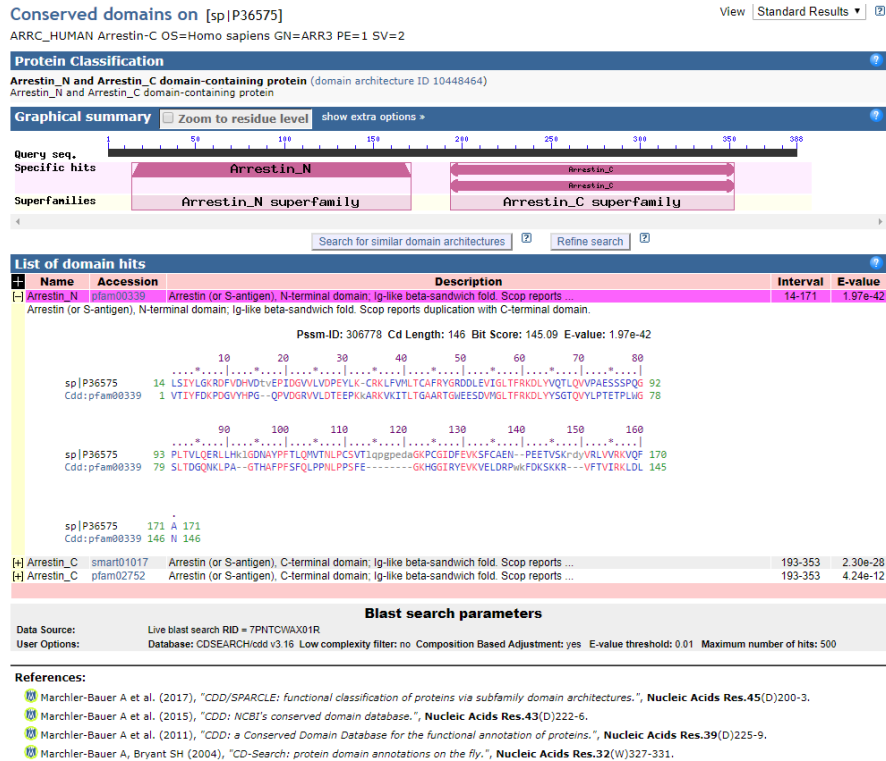


Figure 6. Conserved domains found by BLASTP suite for the human arrestin protein. These results include accession numbers of the domains, the alignment between the query sequence and domain sequence, and the corresponding alignment scores and E-values.

PSI-BLAST

Compared to BLASTP, PSI-BLAST aims to find distant relatives for the query protein, i.e., proteins that are similar to proteins that share similarity with the query sequence. To do that, PSI-BLAST performs the search iteratively. In the first iteration, a list of similar sequences is created using BLASTP. The resulting multiple alignment is used to compute PSSM which is used to substitute the original scoring matrix from the first iteration. This produces a larger and more diverse group of similar proteins. Next, the alignment is run again and the PSSM is recomputed based on the subsequent multiple alignment. The search performed based on PSSM is more sensitive than the search based on the default BLOSUM or PAM matrices since PSSM is based on the multiple alignment that is specific to the query protein. This two-step process can be repeated multiple times.

PSSM is an N by 20 matrix (N is the length of the query sequence) which is calculated from the frequencies of residues at specific positions in the multiple alignment. The scores at the position j in the query sequence (j^{th} row of PSSM) and the amino acid i (i^{th} column of PSSM) are defined as $\log(f_{ij}/q_i)$ where f_{ij} is the frequency of amino acid i at position j in the multiple alignment and q_i is the expected relative frequency of amino acid i . Positive scores mean that a given amino acid is more likely than expected to appear at the specific position. Besides being used for alignment, PSSM is also used to calculate evolutionary conservation of the query sequence (Valdar, 2002)

and to predict protein secondary structure (Jones, 1999; Meng and Kurgan, 2016; Wang et al., 2016), solvent accessibility (Heffernan et al., 2015; Magnan and Baldi, 2014), functional sites (Yan et al., 2016; Zhang and Kurgan, 2017; Zhang et al., 2017), and intrinsic disorder (Disfani et al., 2012; Meng et al., 2017; Mizianty et al., 2010).

PSI-BLAST includes three extra parameters on the top of the parameters that are available for BLAST. They appear below the “Filters and Masking” box (Figure 3). User can provide his/her own PSSM, specify the PSI-BLAST threshold that determines statistical significance of hits that are included in the computation of the PSSM for the next iteration, and define value of the pseudocount parameter. The default value of the threshold is set to 0.005 and we recommended this value as a reasonably good choice. The hits with the E-values that are lower than the threshold are included in calculations for the next iteration. The pseudocount value is used to initialize the frequencies that otherwise would be set to zero. The default value of pseudocount is based on the minimum length description principle (Altschul et al., 2009).

The screenshot shows the BLAST web interface for a PSI-BLAST search. The job title is "sp|P36575|ARRC_HUMAN Arrestin-C OS=Homo sapiens...". The query is "sp|P36575|ARRC_HUMAN Arrestin-C OS=Homo sapiens" with a length of 388 amino acids. The database is "nr" (All non-redundant GenBank CDS). The results table shows the following data:

Description	Max score	Total score	Query cover	E value	Ident	Accession	Select for PSI blast	Used to build PSSM
<input type="checkbox"/> arrestin-C [Homo sapiens]	797	797	100%	0.0	100%	NP_004303.2	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> arrestin-C [Homo sapiens]	796	796	100%	0.0	99%	AAC78395.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> X-arrestin [Homo sapiens]	795	795	100%	0.0	99%	AAB84302.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> arrestin 3, retinal (X-arrestin), isoform CRA_b [Homo sapiens]	795	795	100%	0.0	99%	EAX05349.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> ARR3 [Homo sapiens]	793	793	100%	0.0	99%	CAG33130.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<input type="checkbox"/> PREDICTED: arrestin-C [Pongo abelii]	793	793	100%	0.0	99%	XP_002834739.1	<input checked="" type="checkbox"/>	<input type="checkbox"/>

Figure 7. Output produced by the first iteration of PSI-BLAST for the human arrestin protein. The output is similar to the output from BLASTP except for the Descriptions box where the user can select sequences to generate PSSM for next iteration.

Figure 7 shows the results of the first iteration of PSI-BLAST for the sample query protein, the human arrestin, with the PSI-BLAST threshold = 0.001 and the pseudocount set to the default value. Similar to BLASTP, the corresponding webpage includes the general information

(summary of search parameters) and the Graphic Summary, Descriptions and Alignments boxes. One difference from the output of BLASTP is that the Descriptions box allows for selection of the aligned sequences that will be used to generate PSSM for the next iteration. The aligned sequences that score below the threshold from the previous iteration are marked in yellow. Clicking the “Go” button (middle of Figure 7) results in the computation of the next iteration (iteration 2) of PSI-BLAST. The search typically converges (no new sequences below threshold are found) after several iterations. The search for the human arrestin protein converges after just four iterations. The results for every iteration include the multiple alignment and PSSM; except for the first iteration when PSSM is not yet available. They can be downloaded using a link at the top of the webpage. The PSSM is formatted based on the ASN.1 format from the BLASTP suite that can be read by AsnTool from the NCBI’s toolkit. A plain text PSSM is generated by a standalone BLAST+ program. Correspondingly, we give an example plain-text PSSM in next section that describes BLAST+.

BLAST+ - STANDALONE BLAST

BLAST+ must be run on a local computer. It can be installed on the Windows, Linux and MacOS platforms. The installation package and sources can be downloaded from <ftp://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/LATEST/>. BLAST+ includes BLASTP, BLASTN, BLASTX, TBLASTN, TBLASTX, PSI-BLAST, DELTA-BLAST, and reverse PSI-BLAST (RPS-BLAST) programs. These programs are the same as the programs available via WEB BLAST, except for RPS-BLAST that searches a query sequence against a database of PSSMs. PHI-BLAST from the WEB BLASTP suite is integrated into PSI-BLAST in BLAST+ and can be accessed with the `-phi_pattern` parameter. Similarly, MEGABLAST and discontinuous MEGABLAST from the WEB BLASTN suite are integrated into BLASTN in BLAST+ and can be used with the `-task` parameter.

One advantage of BLAST+ compared to the web-based versions is that users have the choice to use their own databases for the search or to utilize one of dozens of already preprocessed databases that can be downloaded from <ftp://ftp.ncbi.nlm.nih.gov/blast/db/>. The former choice requires users to provide the sequences in a specific format that is compatible with BLAST. The `makeblastdb` program that is part of BLAST+ supports this task. First, the sequences must be prepared in the FASTA format. The `makeblastdb` takes multiple FASTA-formatted files as input and it outputs a properly formatted BLAST database. Here is an example call to run this program:

```
makeblastdb -in 'swiss1.fa swiss2.fa' -dbtype prot -out swiss
```

This above command creates a BLAST database named “swiss” using `swiss1.fa` and `swiss2.fa` FASTA files. The database type (`dbtype`) need be specified as `prot` (for proteins) or `nucl` (for nucleotides).

BLAST+ should be executed from command line. We give an example command to run PSI-BLAST. The query sequence is the same that we use in the WEB BLASTP example, the human arrestin protein, and we use the available SWISS-PROT database as the target database:

```
psiblast -query query.fa -db ./blastdb/swissprot -num_iterations 3 -out result.out -
out_ascii_pssm pssm.out -E-value 0.001
```

The input query.fa (“-query query.fa”) query file is in the FASTA format and includes only the arrestin. We specify the target database as swissprot (“-db ./blastdb/swissprot”). The number of PSI-BLAST iterations is set to 3 (“-num_iterations 3”) and the E-value threshold is specified as 0.001 (“-E-value 0.001”). The results will be saved into the result.out file (“-out_result.out”) and the corresponding PSSM will be stored in the “pssm.out” file (“-out_ascii_pssm pssm.out”). The command line also allows to specify values of other parameters, such as the maximal number of sequences used to generate PSSM, scoring matrix, gap penalty, word size, filters, etc. The default values are used if they are not specified. We recommend the documentation available at <https://www.ncbi.nlm.nih.gov/books/NBK279690/> for further reading on how to setup and use BLAST+.

```
>P15887.1 RecName: Full=S-arrestin; AltName: Full=48 kDa protein; AltName: Full=Retinal
S-antigen; Short=S-AG; AltName: Full=Rod photoreceptor Arrestin
Length=403

Score = 561 bits (1447), Expect = 0.0, Method: Composition-based stats.
Identities = 199/391 (51%), Positives = 277/391 (71%), Gaps = 10/391 (3%)

Query 4 VFKKTSNKGKLSIYLGKRDFVDHVDTVVEPIDGVVLDVPEYLKCRKLFVMLTCAFRYGRDD 63
      +FKK S + ++IYLGKRD++DHV VEP+DGVVLDVPE +K +K++V LTCAFRYG++D
Sbjct 13 IFKKVSRDKSVTIYLGKRDYIDHVSQVEPVDGVVLDVPELVKGGKVVYVTLTCAFRYQGED 72

Query 64 LEVIGLTFRKDLYVQTLQVVPAAESSSPQGPLTVLQERLLHKLKLDNAYPFTLQMVNTLPCS 123
      ++VIGLTFR+DLY +QV P + T LQ LL KLGDN YPF L LPCS
Sbjct 73 IDVIGLTFRRDLYFSRVQVYPPVGA--MSAPTQLQLSLLKLDNAYPFTLQMVNTLPCS 130

Query 124 VTLQPGPEDAGKPCGIDFEVKSFCAE---NPEETVSKRDYVRLVVRKVQFAPPEAGPGPS 180
      V LQP P+D GK CG+DFEVK+F + E+ + K+ VRL++RKVQ APPE GP P
Sbjct 131 VMLQAPAPQDVGKSCGVDFEVKAFATDITDAEEDKIPKKSSVRLIRKVVQHAPPEMGPQPC 190

Query 181 AQTIRRFLLSAQPLQLQAWMDREVHYHGEPISVNVSIINNCTNKVIKKIKISVDQITDVVL 240
      A+ +F +S +PL L + +E+++HGEPI V V++ N T KV+KKIK+SV+QI +VVL
Sbjct 191 AEASWQFFMSDKPLHLSVLSKEIYFHGEPIPVTVTVTNNTKVVKKIKVSVQIANVVL 250

Query 241 YSLDKYTKTVFIQEFTETVAANSSFSQSFVTPILAASCQKRGALDGLKHKHEDTNLASS 300
      YS D Y K V +E E V NS+ +++ + P+LA + ++RG+ALDGLKHKHEDTNLASS
Sbjct 251 YSSDYVVKPVASEETQEKVQPNSTLTKTLVPLANNRERRGIALDGLKHKHEDTNLASS 310

Query 301 TIIRPGMDKELGILVSYKVRVNLVSCGGILGDLTASDVGVLPVLVLIHPK---PSHEA 357
      TII+ G+D+ ++GILVSY ++V L VS G LG+LT+S+V E+P L+HP+ P+ E+
Sbjct 311 TIIKEGIDRTVMGILVSYHIKVKLTVS--GFLGELTSSEVATEVPFRLMHPQPEDPAKES 368

Query 358 ASSEDIVIEEFTRKGEESQKAVEAEGDEGS 388
      E++V EEF R+ +++ + E + DE +
Sbjct 369 VQDENLVFEFARQNLKDTGENTEGKKDEDA 399
```

Figure 8. Results for an example similar sequence generated by the standalone PSI-BLAST for the query human arrestin protein.

The above example PSI-BLAST command generates two files. The first, result.out, lists 129 sequences that are sufficiently similar to our query, i.e., E-value < 0.001. The file provides detailed information including the accession numbers, multitude of scores that quantify quality of the alignment, and the alignment itself. Sample results for one of the hits are shown in Figure 8. The UniProt identifier of the aligned sequence is P15887.1. The alignment score, which is calculated based on PSSM, is 561 and this sequence shares 51% identical residues with the arrestin chain. The alignment, which is shown below these statistics, organizes the sequences into blocks. The first line in each block is the query sequence and the third line is the aligned/hit

sequence. The positions for both sequences are given at the beginning and end of the lines. The sequence in the second line in each block (between the chains of the query and hit) is the match sequence. Letters in the match sequence denote matching residues while “+” means that the residues do not match and that the corresponding score is positive. Moreover, blank space denotes that the score is negative. The second file provides PSSM that is shown in Figure 9. PSSM is formatted as N by 20 matrix where N is the length of the query sequence. We show the first 13 rows that correspond to the first 13 residues in the arrestin sequence. Each row is one position in the query sequence. The log odds scores in a given row are calculated based on the frequency of the 20 amino acids at this position in the multiple alignment generated in the last iteration of PSI-BLAST.

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
1 M	-1	-1	-2	-3	-2	0	-2	-3	-2	1	2	-1	6	0	-3	-2	-1	-2	-1	1
2 S	1	-1	0	-1	-1	0	-1	-1	-1	-2	-2	-1	-1	-3	-1	4	3	-3	-2	-1
3 K	-1	4	-1	-1	-3	1	0	-2	-1	-2	-2	4	-1	-3	-2	-1	0	-3	-2	-1
4 V	-1	-3	-3	-4	-1	-3	-3	-4	-4	3	1	-3	1	-1	-3	-2	-1	-3	-2	4
5 F	-3	-3	-4	-4	-3	-4	-4	-4	-1	-1	0	-4	-1	7	-4	-3	-3	1	4	-1
6 K	-1	2	-1	-1	-4	1	0	-2	-1	-3	-3	6	-2	-4	-2	-1	-1	-4	-3	-3
7 K	-1	2	-1	-1	-4	1	0	-2	-1	-3	-3	6	-2	-4	-2	-1	-1	-4	-3	-3
8 T	1	-2	-1	-2	3	-1	-2	-2	-2	0	-1	-1	-1	-2	-2	2	4	-3	-2	0
9 S	1	-1	0	-1	2	-1	-1	-1	-2	-2	-3	-1	-2	-3	-1	4	2	-3	-2	-2
10 S	-1	2	-1	-2	-3	-1	-1	-1	-2	-3	-3	0	-3	-4	6	1	-1	-4	-3	-3
11 N	-2	-1	6	4	-4	-1	0	-1	0	-4	-4	-1	-3	-4	-2	0	-1	-5	-3	-4
12 G	-1	-1	1	-1	2	-1	-1	5	-2	-4	-4	2	-3	-4	-2	-1	-2	-3	-3	-3
13 K	0	1	-1	-1	-3	0	0	-2	-1	-2	-2	5	2	-3	-2	1	-1	-3	-2	-2

.....

Figure 9. PSSM generated by the standalone PSI-BLAST based on the query for the human arrestin protein. We shows values for the first 13 residues in the arresting sequences. The first two columns are the sequence position and one code amino acid type at this position. The following 20 columns are the scores for every amino acid type at a given position.

BLASTN SUITE - BLAST AGAINST NUCLEOTIDE DATABASES

BLASTN suite searches nucleotide sequence query against a nucleotide database. It includes three programs: MEGABLAST, discontinuous MEGABLAST and BLASTN. MEGABLAST was designed for searching highly similar sequences, such as in case of the intra-species comparisons (Morgulis et al., 2008). Discontinuous MEGABLAST should be used to search more dissimilar sequences, which are typical for cross-species comparisons. Finally, BLASTN is suitable for short queries and cross-species comparisons.

The webpage of the BLASTN suite is shown in Figure 10. Similar to the BLASTP suite, it has three parts:

- (1) “Enter Query Sequence” box should be used to provide the query sequences in one of the two ways: directly enter query sequence into the text box or upload it in a file. Besides the FASTA-formatted sequences, users can provide database accession number or NCBI gi number. Also, a segment of the query sequence that can be specified in “Query subrange” box may be used for the search. Like for BLASTP, BLASTN suite also allows

aligning sequences provided by the users. To do so, users should check the “Align two or more sequences” checkbox and input their own sequences for the alignment.

(2) “Choose Search Set” box provides selection of the target database or organism. The search can be restricted to a subset of selected database using “Organism” “Exclude” “Limit to” and “Entrez Query” options.

(3) “Program Selection” box should be used to select one of the three program: MEGABLAST, discontinuous MEGABLAST and BLASTN.

The “Algorithm parameters” link at the bottom of the page opens additional menu where users can choose the scoring matrix and can set values of the gap penalties and other search parameters.

The screenshot shows the BLASTN web interface. At the top, there's a navigation bar with "BLAST" and "blastn suite" on the left, and "Home", "Recent Results", "Saved Strategies", and "Help" on the right. Below this is the "Standard Nucleotide BLAST" header. The main form is divided into several sections: "Enter Query Sequence" with a text area containing a FASTA sequence and a "Job Title" field; "Choose Search Set" with radio buttons for "Human genomic + transcript", "Mouse genomic + transcript", and "Others (nr etc.)", and a dropdown menu for "Nucleotide collection (nr/nt)"; "Program Selection" with radio buttons for "Highly similar sequences (megablast)", "More dissimilar sequences (discontiguous megablast)", and "Somewhat similar sequences (blastn)"; and a "BLAST" button at the bottom left. There is also an "Algorithm parameters" link at the bottom right.

Figure 10. Web interface for the BLASTN suite.

We run an example query for the retina glycoprotein gene from *Drosophila Melanogaster* using the nr database and BLASTN. We use the default settings for the algorithm. The output of BLASTN is formatted the same way as the output of BLASTP. At the top of the page it gives general information (Figure 11) that includes the target database name, query sequence identifier

and length, type and description of the query molecule as well as details of the BLASTN parameters. This is followed by:

- (1) “Graphic Summary” box shown in Figure 12 that provides a graphical representation of the alignments of the 79 sequence hits. Each hit is shown as a colored bar where the color represents the alignment score. Detailed information for a specific hit will be shown when the corresponding bar is clicked. It includes description of the hit sequence, alignment score and E-value. The alignment scores quantify the degree of similarity while the E-values quantify number of hits one can expect to observe by chance.
- (2) “Descriptions” box shown in Figure 13 provides detailed statistics and database accession identifiers for each hit. The hits are sorted by E-value in the ascending order. Besides the E-values and alignment scores, this box also gives the maximal score, query coverage and identity. The maximal score is the highest alignment score among the aligned segments between the query and hit sequences. The query coverage is the fraction of the query sequence length that is covered by the aligned segments. The identity gives the % of the aligned segments where the two sequences match. The best hit with the 100% query coverage and 100% identity in Figure 13 is the exact same sequences as our query sequence. The following hits are the sequences of the chromosome 3R from *Drosophila Melanogaster* which localize our query.
- (3) “Alignments” box gives detailed alignments along the query sequence. Figure 14 shows an example alignment for one of the hits for the retina glycoprotein gene query. This alignment contains one segment in the 7 to 242 range of the query sequence. Vertical lines between query sequence (first line in each alignment block) and hit sequence (third/subject line in each alignment block) correspond to matched identical nucleotides. In this example there are 197 identities and one gap in the middle of the last block.

BLAST [®] » blastn suite » RID-83YANBCM014 [Home](#) [Recent Results](#) [Saved Strategies](#) [Help](#)

BLAST Results

[Edit and Resubmit](#) [Save Search Strategies](#) [▶ Formatting options](#) [▶ Download](#) [YouTube](#) [How to read this page](#) [Blast report description](#)

Job title: K03274.1 D.melanogaster retina glycoprotein...

RID 83YANBCM014 (Expires on 02-14 03:30 am)	Database Name nr
Query ID Id Query_59931	Description Nucleotide collection (nt)
Description K03274.1 D.melanogaster retina glycoprotein (antigen 24B10) gene, 5' end	Program BLASTN 2.8.0+ ▶ Citation
Molecule type nucleic acid	
Query Length 240	

Other reports: [▶ Search Summary](#) [\[Taxonomy reports\]](#) [\[Distance tree of results\]](#) [\[MSA viewer\]](#)

[+ Graphic Summary](#)
[+ Descriptions](#)
[+ Alignments](#)

Figure 11. Summary information that is shown at the top of the page with the results generated by the BLASTN suite for the retina glycoprotein gene.

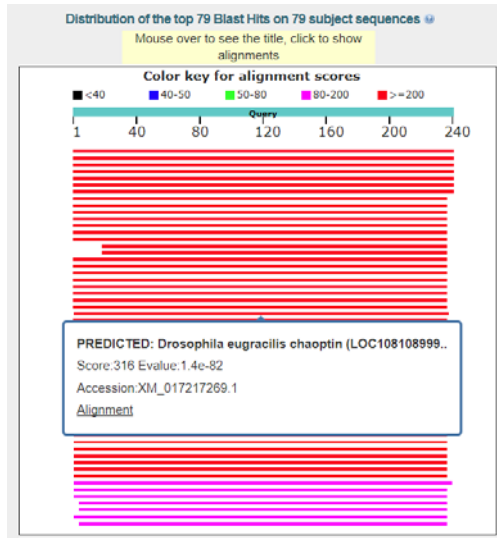


Figure 12. Graphic summary box generated by the BLASTN suite for the retina glycoprotein gene.

Sequences producing significant alignments:

Select: [All](#) [None](#) Selected:0

[Alignments](#) [Download](#) [GenBank](#) [Graphics](#) [Distance tree of results](#)

Description	Max score	Total score	Query cover	E value	Ident	Accession
<i>D.melanogaster</i> .retina glycoprotein (antigen 24B10).gene_5' end	434	434	100%	7e-118	100%	K03275.1
<i>Drosophila melanogaster</i> strain rover (forR) chromosome 3R	426	426	100%	1e-115	99%	GM23396.1
<i>Drosophila melanogaster</i> chromosome 3R	426	426	100%	1e-115	99%	AE014297.3
<i>Drosophila melanogaster</i> clone BACR30B13..complete sequence	426	426	100%	1e-115	99%	AC008221.5
<i>Drosophila melanogaster</i> .chromosome 3R..region 100B-100C..BAC clone BACR11C03..complete sequence	426	426	100%	1e-115	99%	AC008220.5
<i>Drosophila melanogaster</i> strain siltter (fors) chromosome 3R	425	425	100%	4e-115	99%	CP023330.1
<i>Drosophila melanogaster</i> chaoptin gene..complete cds	423	423	100%	1e-114	99%	AH000987.2

Figure 13. Part of the description box output by the BLASTN suite for the retina glycoprotein gene.

[Download](#) [GenBank](#) [Graphics](#) [Next](#) [Previous](#) [Descriptions](#)

Drosophila grimshawi GH14339 (DgriGH14339), mRNA
 Sequence ID: [XM_001996113.1](#) Length: 3948 Number of Matches: 1

Range 1: 7 to 242 [GenBank](#) [Graphics](#) [Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
246 bits(272)	2e-61	197/236(83%)	1/236(0%)	Plus/Plus
Query 2	CTCGAATTTTCTTTAAGTTCGGCTATGCCTTCTTGACTATAACGCTCATGATCATGATC	61		
Sbjct 7	CTGGAGTTCCTTTAAATTTGGCTATGCCTTCTTCCACATAACACTGATTATAATGATC	66		
Query 62	TGGATGTCGCTGGCTCGCGCCTCCATGTTTCGATCGCGAGATGGAGGAGACGCAATTACCCG	121		
Sbjct 67	TGGATGTCGTTGGCGCGTGCCTCGATGTTTCAGTCGGGAAATGGAGGAGACACACTATCCG	126		
Query 122	CCCTGCACCTACAACGTGATGTGCACCTGCTCCAAGTCTCCACGGATCTGGGGATAGTG	181		
Sbjct 127	CCATGCACCTACAATGTGATGTGCACGTGTTTCAAGGCTCCACAGATCTGGGCATTGTC	186		
Query 182	CACTGCAAGAATGTTCCGTTT-CGGCACTGCCGCGCATGGTGAACCAAGTCAAAGGT	236		
Sbjct 187	CATTGCAAGAATGTTCCGTTTCCAGCACTGCCGCGCATGGTTAACCAATCAAAGGT	242		

Related Information
[Gene-associated gene details](#)

Figure 14. An example alignment produced by the BLASTN suite for the retina glycoprotein gene.

RECENT ADVANCES

Recent developments in high throughput sequencing technologies resulted in the generation of massive datasets and a rapid growth of the sequence databases. For instance, RefSeq has grown from one million records in 2003, to 10 million in 2009 and 150 million in 2018 (<https://www.ncbi.nlm.nih.gov/refseq/statistics/>). Corresponding, searching for similar sequences becomes more time-consuming. Several tools that are faster than the popular BLAST were developed in recent years to address this issue. They include UBLAST (Edgar, 2010), RAPSearch2 (Zhao et al., 2012), Diamond (Buchfink et al., 2015), MMSeq (Hauser et al., 2016), and MMseqs2 (Steinegger and Soding, 2017). Here we focus on the most recent MMseqs2 algorithm.

MMseqs2 (Many-against-Many sequence searching) is specifically designed to search and cluster huge protein sequence datasets. According to published empirical results MMseqs2 is 10,000 times faster than BLAST, and at 100 times speedup it achieves the same sensitivity as BLAST (Steinegger and Soding, 2017). MMSeq2 software can be freely downloaded from <https://github.com/soedinglab/MMseqs2>. Users must prepare their database before searching with MMSeq2. Here, we use SWISS-PROT as the target database, like we did with BLAST. We downloaded the SWISS-PROT sequences and generated the corresponding sequence database for MMseqs2 as follows:

```
mmseqs createdb swissprot.fasta swissprot
```

The swissprot.fasta file includes the FASTA-formatted SWISS-PROT sequences. The above command generates the MMseqs2 database named swissprot. Unlike for BLAST+, the query sequences must be also converted into the MMseqs2's database format. The corresponding command follows:

```
mmseqs createdb arrestin.fasta arrestin
```

The arrestin.fasta file includes the sequence of the human arrestin proteins that we also used with BLAST. MMseqs2 can also accept multiple sequences as the input.

MMseqs2 includes five workflows:

- (1) mmseqs search for searching query sequences against the target database
- (2) mmseqs cluster for clustering sequences by similarity
- (3) mmseqs linclust which is a less sensitive but faster program for clustering sequences by similarity
- (4) mmseqs clusterupdate for incremental updating of an existing clustering
- (5) mmseqs taxonomy for assignment of the query chain to a taxonomy by computing the lowest common ancestor

We focus our example on the mmseqs search that is equivalent to the BLASTP search:

```
mmseqs search arrestin swissprot result.out tmp -e 0.0005 -a
```

The human arrestin is our query, SWISS-PROT is the target database, and the search results are saved in the result.out file. The “tmp” is directory where temporary files are saved and “-e”

parameter should be used to specify the threshold of the E-values; for our example only the sequences with E-value < 0.0005 will be listed. Finally, the “-a” option specifies that alignment backtraces should be used. Total of 40 hits are found in SWISS-PROT for the human arrestin. The output result.out file can be converted into BLAST output format using the following command:

```
mmseqs convertalis arrestin swissprot result.out result.m8
```

The BLAST output-formatted results are saved in the results.m8 file (Figure 15). This text file is formatted as a tab-separated list with 12 columns: (1) query id, (2) hit accession id, (3) identity between query and hit sequences, (4) the length of alignment, (5) number of mismatches, (6) opening gap number, (7-10) the start position and end position of alignment in query and hit sequence, (11) E-value, and (12) alignment bit score. Using the same command and adding option “-format-mode 1” gives the alignment of the query sequence and hit sequences in the FASTA format. Figure 16 shows the alignment between the query chain and the sequence of the Q5DRQ4.1 protein, which is the second hit in Figure 15. The first line in Figure 16 gives the same information and statistics about the alignment that are given in Figure 15 while the second line and third lines give the aligned query and hit sequences, respectively.

Query_id	Hit_id	Identity	Alignment_length	Mismatches_num	opening_gap_num	query_start_position	query_end_position	Hit_start_position	Hit_end_position	E-value	Bit_score
P36575	P36575.2	1.000	388	0	0	1	388	1	388	7.03E-253	774
P36575	Q5DRQ4.1	0.879	389	45	1	2	388	5	393	3.89E-221	683
P36575	Q7YS78.2	0.867	391	49	1	1	388	1	391	3.20E-219	677
P36575	Q9N0H5.2	0.843	389	60	1	1	388	1	389	7.98E-211	653
P36575	Q9EQP6.1	0.841	372	55	1	1	368	1	372	6.80E-200	621
P36575	P51482.1	0.650	369	128	1	2	370	5	372	3.98E-161	509
P36575	P51481.1	0.644	369	130	1	2	370	5	372	4.52E-160	506
P36575	P51483.1	0.628	369	136	1	2	370	5	372	1.47E-152	484
P36575	P17870.1	0.588	389	139	2	2	370	6	393	8.46E-148	471
P36575	P49407.2	0.586	389	140	2	2	370	6	393	1.69E-147	469
.....											

Figure 15. The MMseqs2 output for the human arrestin search against the SWISS-PROT database. We show the 10 hits with the lowest E-values from the complete set of 40 hits.

```
>P36575 Q5DRQ4.1          0.879  389  45  1  2  388  5  393
3.89E-221          683
SKVFFKTSSNGKLSIYLKGRDFVDHVDVTEPIDGVVLDPEYLGKCRKLFVMLTCAFYGRDDLEVI GLTFRKDLVYVLTQVVP AES
SSPQGPLTVLQERLLHKLGDNAYPFTLQMVNLPCSVTLQPGPEDAGKPCGIDFEVKSFCFAENPEETVSKRDYVRLVVRKVQFAPP
EAGPGPSAQTIRRFLLSAQPLQLQAWMDREVHYHGEPISVNVSIINNCTNKVIKKIKISVDQITDVLVYSLDKYTKTVFIQEFTEITV
AANSSFSQSFVTPILAASCQKRGLALDGKLGKHEDTNLASSTIIRPGMDKELLGILVSYKVRVNLVMSVCGGILGDLTASDVGVGVELP
LVLIHPKPSHEAASSEDIVIEEFTRK--GEEESQKAVEAEGDEGS
SKVFFKTSSNGKLSIYLKGRDFMDHVDVTEPIDGVVLDPEYLGKGRKMFVILTCAFYGRDDLDVI GLTFRKDLVYVLTQVVP AES
NSPQGPLTVLQERLLHKLGENAYPFTLQMVANLPCSVTLQPGPEDSGKACGVDFEVKSFCFAENLEEKVSKRDSVRLVVRKVQFAPM
EPGPGPWAQTIRRFLLSVQPLQLQAWMDKEVHYHGEPISVNVSIINNSTSKVIKKIKISVDQITDVLVYSLDKYTKTVFIQEFTEITI
AANSSFTQSFVTPLLSANCRQGLALDGKLGKHEDTNLASSTIVRPGMNKELLGILVSYKVRVNLVMSVCGGILGDLTASDVGVGVELP
LTLIHPKPSQETTSSSEDIVIEEFARQEDGGEKQKALAEEGDEGS
```

Figure 16. Alignment of query sequence and Q5DRQ4.1 in FASTA-format for the human arresting query that are produced by MMSeq2.

Users who want faster (less sensitive) searches can adjust sensitivity of MMseq2 with the “-s” parameter. The sensitivity parameter can be set to 1.0 (much faster), 4.0 (fast), 5.7 (default) and 7.5 (slower and more sensitive). MMseqs2 also offers iterative searching similar to PSI-BLAST in order to satisfy the requests for more sensitive searches. The command for the iterative search is the same as a regular search with the addition of the “--num-iterations” parameter. Like PSI-

BLAST, MMseqs2 also generates PSSM for the query sequence. The following commands should be used to obtain PSSM for the human arrestin:

```
mmseqs search arrestin swissprot result.out tmp -e 0.0005 -a --num-iterations 5
mmseqs result2profile arrestin swissprot result.out result.profile
mmseqs profile2pssm result.profile result.pssm
```

The first command produces results for the MMSeq search with 5 iterations. The second command converts the search results into a profile format that is stored in the result.profile file. The third command converts this profile into PSSM that is saved in the result.pssm file. The multiple alignment between the query and hit sequences can be generated with the result2msa command from the generated above result.out file as follows:

```
mmseqs result2msa arrestin swissprot result.out result.msa
```

The result.msa file stores the multiple alignments.

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Describes the BLAST+ suite

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Introduces the MMseqs2 algorithm

INTERNET RESOURCES

<ftp://ftp.ncbi.nlm.nih.gov/blast/executables/blast+/LATEST/>
BLAST+

<ftp://ftp.ncbi.nlm.nih.gov/blast/db/>
BLAST databases

<https://github.com/soedinglab/MMseqs2>
MMSeq2

<https://blast.ncbi.nlm.nih.gov/Blast.cgi>
WEB BLAST